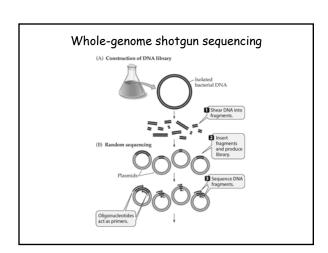
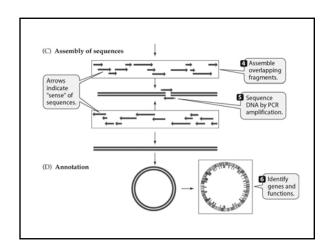
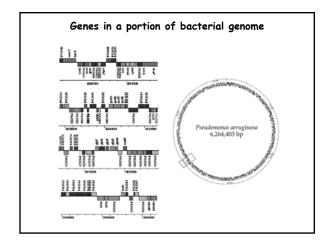
# Microbial Genomics and Chromosome Organization

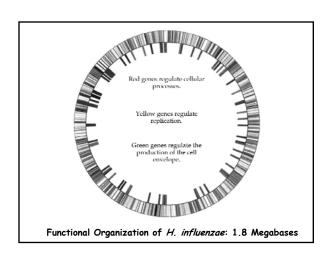
- Microbial Chromosome Organization
- Generation of full genome sequences
- Genomic Structure & Functional Genomics
- Genome size vs. No. of orfs
- Minimal genome concept
- $\bullet$  Lessons from full genomes

Organism	Element	Description	Chromosomal &
Prokaryote	Chromosome	Extremely long, usually circular, double-stranded DNA molecule	nonchromosoma genetic element
	Plasmid	Typically a relatively short, usually circular, double- stranded DNA molecule, which is extrachromosomal	generic element
Eukaryote	Chromosome	Extremely long, linear, double- stranded DNA molecule	
	Plasmid <sup>d</sup>	Typically a relatively short circular or linear double- stranded DNA molecule, which is extrachromosomal	
All Organisms	Transposable elements	Double-stranded DNA molecule always found within another DNA molecule	
Mitochondrion or chloroplast	Chromosome	Intermediate-length DNA molecules, usually circular	
Virus	Genome	Single- or double-stranded DNA or RNA molecule	

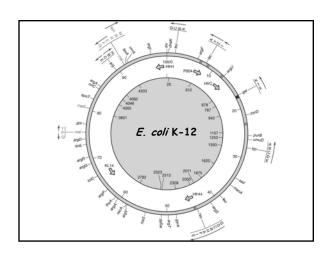








Comparison of regulatory genes in select bacterial genomes					
Microorganism	# Genes in the Genome	# Regulatory Proteins	% of Total		
Pseudomonas aeruginosa	5570	468	8.4		
Escherichia coli	4289	250	5.8		
Bacillus subtilis	4100	217	5.3		
Mycobacterium tuberculosis	3918	117	3.0		
Helicobacter pylori	1566	18	1.1		



## E. coli K-12 Genome Organization Lessons

Cured of F-plasmid and Lambda phage: 4.6 Mbs

88% of genome comprised in 4200 orfs 1% tRNAs and rRNAs

0.5% noncoding repetitive sequences
10% regulatory sequences, including promotors,
operators, origin and terminus of DNA replication

70% single copy genes Some gene clusters like lac and trp operons ~6% polycistronic mRNAs

## E. coli K-12 Genome Organization Lessons

Gene orientation can be in both directions, however, highly expressed genes in the same direction as DNA replication forks travel.

For example, all 7 rrn operons Rem: transcription always 5' to 3'

Also found were several different cryptic, defective prophages and IS elements.

18% of genome from horizontal gene transfer, including large regions (~40Kb) known as islands of pathogenicity.

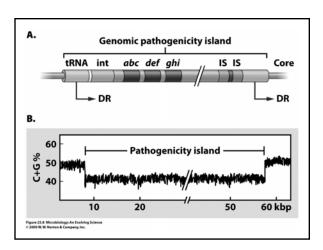
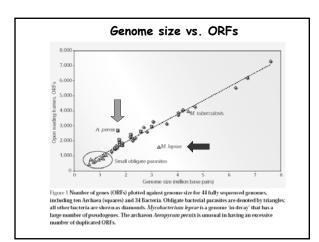


Table 15.2	Gene function	in bacterial gen	omes			
	Percentage of genes on chromosome in that category					
Functional categories	Escherichia coli (4.64 Mbp) <sup>a</sup>	Haemophilus influenzae (1.83 Mbp) <sup>a</sup>	Mycoplasma genitalium (0.58 Mbp) <sup>a</sup>			
Metabolism	21.0	19.0	14.6			
Structural	5.5	4.7	3.6			
Transport	10.0	7.0	7.3			
Regulation	8.5	6.6	6.0			
Translation	4.5	8.0	21.6			
Transcription	1.3	1.5	2.6			
Replication	2.7	4.9	6.8			
Other, known	8.5	5.2	5.8			
Unknown	38.1	43.0	32.0			



	Tred by	5 TOTAL	E CENT	Streets Streets	nerje adire	Herit
Organism (number of genes)	Chlon	Trichland	Prings.	Prairies &	6-April 1900 SA	Ancestral stock
Mycoplasma genitalium (470)	+	-	-	-	-	Bacillus-Clostridium
Buchnera species (588)	+	-	+	+	+	Gamma- proteobacteria
Rickettsia prowazekii (834)	-		-	-	-	Alpha- proteobacteria
Chlamydia trachomatis (894)	+	-	+	-	-	Main line
Treponema pallidum (1,041)	+	-	-	-	-	Main line
Mycobacterium leprae (1.604)	Partial	In decay	+	+	+	Bacillus-Clostridiun

Figure 2 Many routes to intracellular adaptation. The differing presence (+) or absence (-) of certain metabolic pathways in the streamlined genomes of parasitic bacteria shows how variable the process may be.

#### Global Transposon Mutagenesis and a Minimal Mycoplasma Genome

Clyde A. Hutchison III,<sup>1,2+</sup> Scott N. Peterson,<sup>1+</sup>† Steven R. Gill,<sup>1</sup> Robin T. Cline,<sup>1</sup> Owen White,<sup>1</sup> Claire M. Fraser,<sup>1</sup> Hamilton O. Smith,<sup>1</sup>‡, J. Craig Venter<sup>1</sup>‡§

Hamilton O. Smith, '\$ J. Craig Venter' \{\gamma}\\$
Mycoplasma genitalium with 517 genes has the smallest gene complement of any independently replicating cell so far identified. Global transposon mutagenesis was used to identify nonessential genes in an effort to learn whether the naturally occurring gene complement is a true minimal genome under laboratory growth conditions. The positions of 2209 transposon insertions in the completely sequenced genomes of M. genitalium and its Close relative M. pneumoniae were determined by sequencing across the junction of the transposon and the genomic DNA. These junctions defined 1354 distinct sites of insertion that were not lethal. The analysis suggests that 265 to 350 of the 480 protein-coding genes of M. genitalium are essential under laboratory growth conditions, including about 100 genes of unknown function.

265 to 350 genes are the minimum necessary genome

# The complete genome of the hyperthermophilic bacterium Aquifex aeolicus

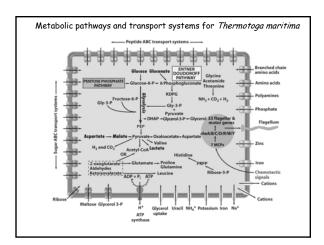
Gerard Deckert\*\*, Patrick V. Warren\*\*, Terry Gaasterland\*, William G. Young\*, Anna L. Lenox\*, David E. Graha Ross Overboek; Marjory A. Snead\*, Martin Keller\*, Monette Aujay\*, Robert Hubert, Robert A. Feldman\*, Jay M. Short, Gay J. Olsen, S. Ronald V. Swansson\*, 1864 Swansa Miller Ross San Bigs, California 9212, USA

\*\*Illinearis Graphanian, 1866 Swansa Miller, Brast San Bigs, California 9212, USA

\*\*Illinearis\* and Carparation, 1866 Swansa Miller, Augus National Labournis\*, Augus, 1858 Biolis 6619, USA

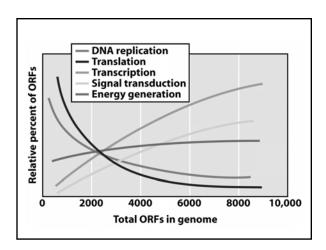
\*\*Shyenment of Marchidge, University of Biotics, USAs, Illinea 6100, USA

Aquillor accidicus was one of the earliest diverging, and is one of the most thermophilic, bacteris known. It can grow on hydrogen, oxygen, carbon dioxide, and mineral salts. The complex metabole machinery needed for A. accidicus to function as a chemolithous orboroph jan organism which uses an inorganic charbon source for bio synthesis and an inorganic chemical energy source) is encoded within a genome that is only one-third the size of the E. col genome. Metabolic flexibility seems to be reduced as a result of the limited genome size. The use of oxygen judicel at very low concentrations) as an electron acceptor is allowed by the presence of a complex respiratory apparatus. Although this organism goves at 95°C, the extreme thermal limit of the Eacteria, oxyl are separation for thempolity are apparent from the genome. Here we describe the complete genome sequence of 1,551,335 base pairs of this evolutionarity and physiologically interesting organism.



	tion of genes of I genomes (Part		wn functio	n among	selected		
Organism	Genome Size (Mbp)		of ORFs oding)		nown	Uniqu	ie ORF
Aeropyrum pernix K1	1.67	1,885	(89%)				
A. aeolicus VF5	1.50	1,749	(93%)	663	(44%)	407	(27%
A. fulgidus	2.18	2,437	(92%)	1,315	(54%)	641	(26%
B. subtilis	4.20	4,779	(87%)	1,722	(42%)	1,053	(26%
B. burgdorferi	1.44	1,738	(88%)	1,132	(65%)	682	(39%
Chlamydia pneumoniae AR39	1.23	1,134	(90%)	543	(48%)	262	(23%
Chlamydia trachomatis MoP <sub>n</sub>	1.07	936	(91%)	353	(38%)	77	(8%
C. trachomatis serovar D	1.04	928	(92%)	290	(32%)	255	(29%
Deinococcus radiodurans	3.28	3,187	(91%)	1,715	(54%)	1,001	(31%
E. coli K-12-MG1655	4.60	5,295	(88%)	1,632	(38%)	1,114	(26%
H. influenzae	1.83	1,738	(88%)	595	(35%)	237	(14%
H. pylori 26695	1.66	1,589	(91%)	744	(45%)	539	(33%
Methanobacterium thermotautotrophicum	1.75	2,008	(90%)	1,010	(54%)	496	(27%

	Genome Size	No.	of ORFs	Hek	nown		
Organism	(Mbp)		oding)		ction	Uniqu	ue ORF:
Methanococcus jannaschii	1.66	1,783	(87%)	1,076	(62%)	525	(30%)
M. tuberculosis CSU#93	4.41	4,275	(92%)	1,521	(39%)	606	(15%)
M. genitalium	0.58	483	(91%)	173	(37%)	7	(2%)
M. pneumoniae	0.81	680	(89%)	248	(37%)	67	(10%
N. meningitidis MC58	2.24	2,155	(83%)	856	(40%)	517	(24%)
Pyrococcus horikoshii OT3	1.74	1,994	(91%)	589	(42%)	453	(22%)
Rickettsia prowazekii Madrid E	1.11	878	(75%)	311	(37%)	209	(25%)
Synechocystis sp.	3.57	4,003	(87%)	2,384	(75%)	1,426	(45%)
T. maritma MSB8	1.86	1,879	(95%)	863	(46%)	373	(26%)
T. pallidum	1.14	1,039	(93%)	461	(44%)	280	(27%
Vibrio cholerae El Tor N1696	4.03	3,890	(88%)	1,806	(46%)	934	(24%)
Totals:	50.60	52,462	(89%)	22.3	58 (43%)	12,161	(23%)



#### Lessons from full genomes

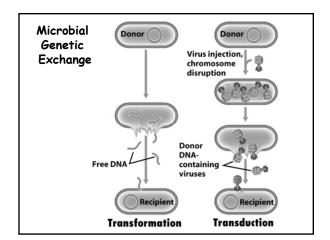
- Size range 600Kb to 12Mb
- Vast number of putative genes with no known function
- Pathogenicity conferred by "Pathogenicity Islands" 44.5Kb in *Bacillus anthracis*
- Symbiotic Island of >600Kb in *Sinorhizobium loti* including genes for nodulation and N-fixation
- Adaptive gene losses in parasitic bacteria
   Rickettsia and Chlamidia are ATP thieves using
   the same "alien" ADP/ATP translocase
- Relative proportions of functional genes

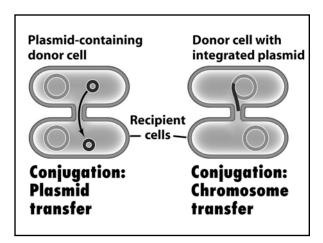
#### Microbial Genetic Exchange and Plasmids

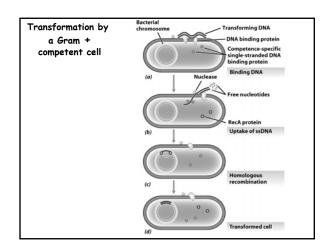
• Microbial Genetic Exchange is unidirectional!

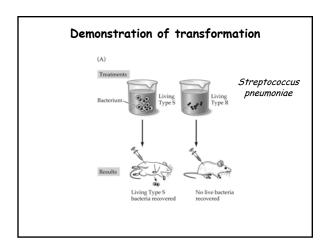
Transformation Transduction Conjugation

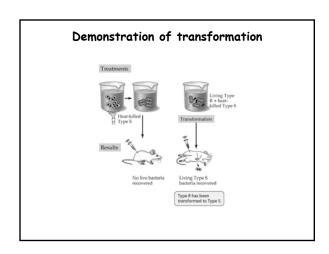
- Each requires **Homologous Recombination**
- Types of plasmids

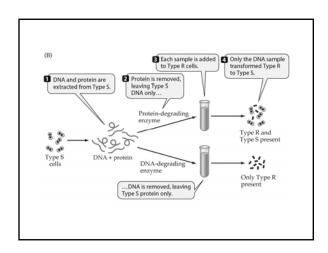


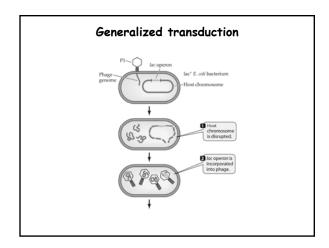


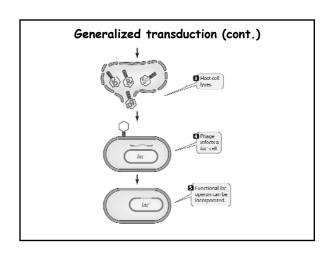


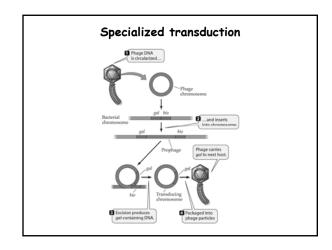


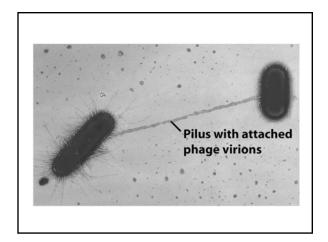


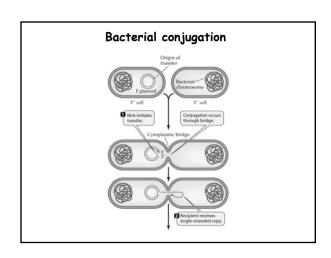


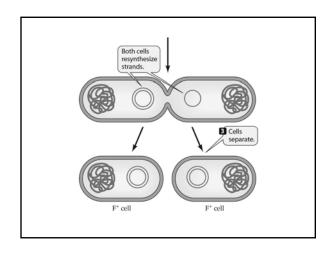


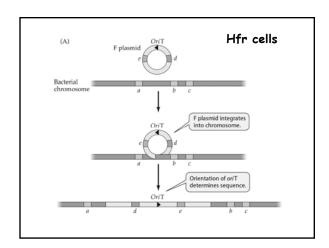


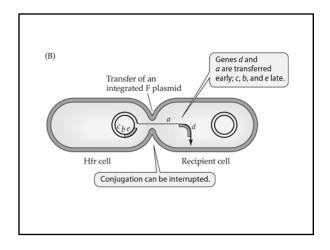


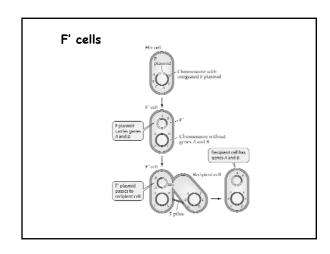


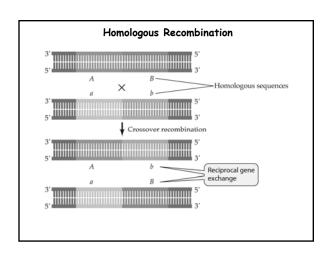


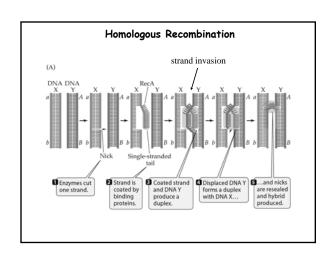


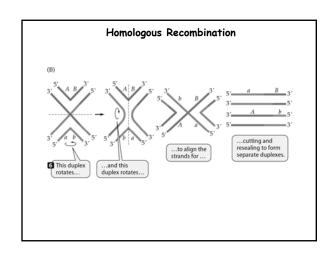


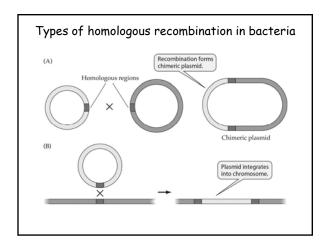


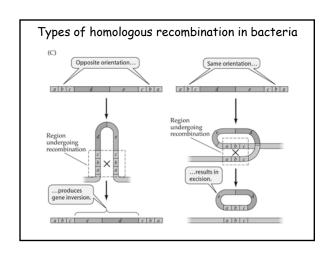


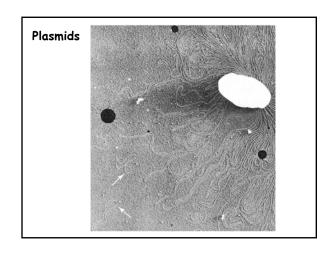


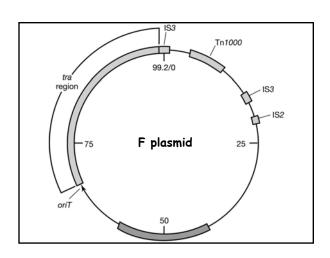


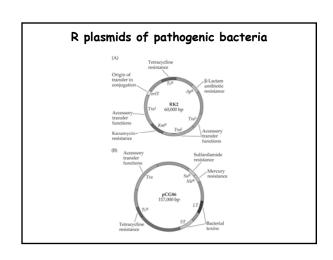


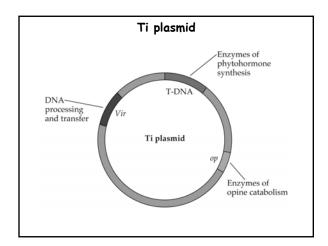












Phenotype class <sup>a</sup>	Organisms <sup>b</sup>
Antibiotic production	Streptomyces
Conjugation	Escherichia, Pseudomonas, Rhizobium, Staphylococcus, Streptococcus, Sulfolobus, Vibrio
Physiological functions Degradation of ectane, camphor, naphthalene Degradation of herbicides Formation of acetone and butanol (○□○ Section 12.20) Lactoee, sucrose or urea utilization and nitrogen fixation Nodulation and symbiotic nitrogen fixation (○□○ Section 19.22) Paremet production	Porudomonas Akadigenes Clotrálium Enteric Posteria Rhizbitum Erubria, Staphylococcus
Resistance Antibiotic resistance ( ○○ Section 20.12) Resistance to cadmium, cobalt, mercury, nickel, and/or zinc ( ○○ Section 19.16) Bacteriocin resistance (and production)	Campylobacter, Enteric bacteria, Neisseria, Staphylococcus Acidocella, Alcaligenes, Listeria, Pendomonas, Staphylococcus Bacillus, Enteric bacteria, Lactococcus, Propionibacterium
Virulence Host cell invasion Coagulase, hemolysin, enterotoxin (ॐ Sections 21.9 and 21.11) Enterotoxin, K antigen (ॐ Sections 12.11 and 21.11) Tumorigenicity in plants (ॐ Section 19.21)	Salmonella, Shigella, Yersinia Staphylococcus Exherichia Agrobacteriam