

Microbial Genomics and Chromosome Organization

- Prokaryotic Chromosome Organization
- Generation of full genome sequences
- Genomic Structure & Functional Genomics
- Genome size vs. No. of orfs
- Minimal genome concept
- Lessons from full genomes

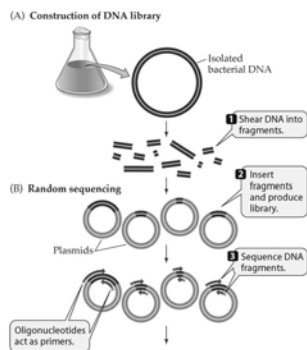
Table 7.1 Kinds of genetic elements

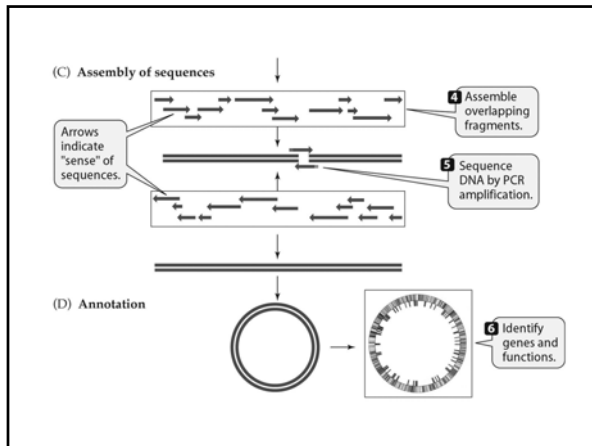
Organism	Element	Description
Prokaryote	Chromosome	Extremely long, usually circular, double-stranded DNA molecule
	Plasmid	Typically a relatively short, usually circular, double-stranded DNA molecule, which is extrachromosomal
Eukaryote	Chromosome	Extremely long, linear, double-stranded DNA molecule
	Plasmid*	Typically a relatively short circular or linear double-stranded DNA molecule, which is extrachromosomal
All Organisms	Transposable elements	Double-stranded DNA molecule always found within another DNA molecule
Mitochondrion or chloroplast	Chromosome	Intermediate-length DNA molecules, usually circular
Virus	Genome	Single- or double-stranded DNA or RNA molecule

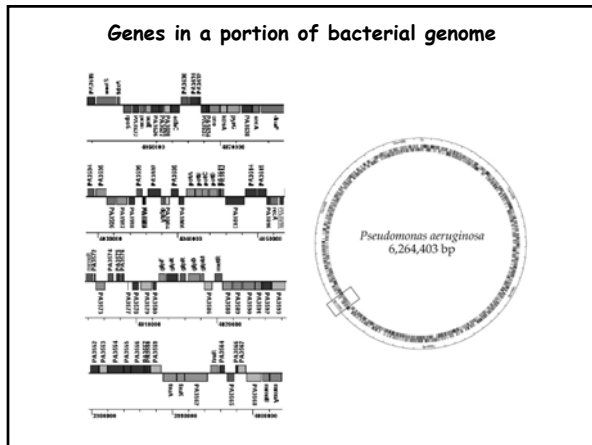
*Plasmids are uncommon in eukaryotes.

Chromosomal & nonchromosomal genetic elements

Whole-genome shotgun sequencing







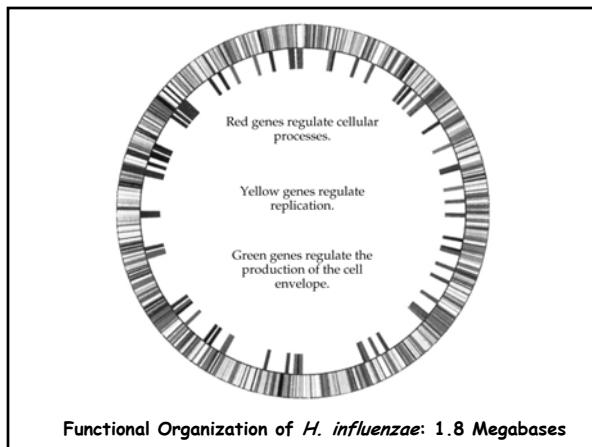
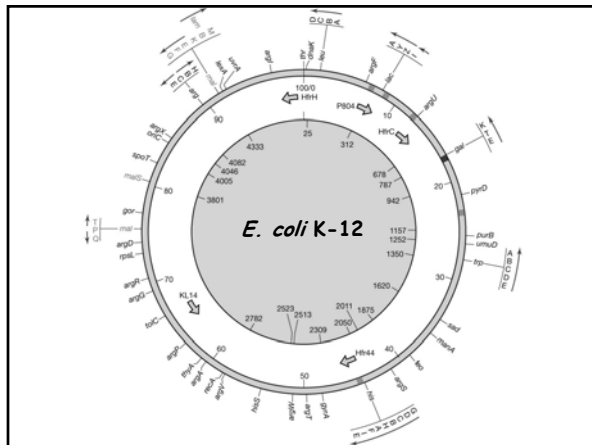


Table 16.2 Comparison of regulatory genes in selected bacterial genomes

Microorganism	# Genes in the Genome	# Regulatory Proteins	% of Total
<i>Pseudomonas aeruginosa</i>	5570	468	8.4
<i>Escherichia coli</i>	4289	250	5.8
<i>Bacillus subtilis</i>	4100	217	5.3
<i>Mycobacterium tuberculosis</i>	3918	117	3.0
<i>Helicobacter pylori</i>	1566	18	1.1



***E. coli* K-12 Genome Organization Lessons**

Cured of F-plasmid and Lambda phage: **4.6 Mbs**

- 88%** of genome comprised in 4200 orfs
- 1%** tRNAs and rRNAs
- 0.5%** noncoding repetitive sequences
- 10%** regulatory sequences, including promoters, operators, origin and terminus of DNA replication

70% single copy genes
 Some gene clusters, e.g., lac, trp, his, and NOT arg
 ~**6%** polycistronic mRNAs

E. coli K-12 Genome Organization Lessons

Gene orientation can be in both directions, however, highly expressed genes in the same direction as DNA replication forks travel.

For example, all 7 *rrn* operons
Rem: transcription always 5' to 3'

Also found were several different cryptic, defective prophages and IS elements.
18% of genome from horizontal gene transfer, including large regions (~40Kb) known as **islands of pathogenicity**.

Table 15.2 Gene function in bacterial genomes

Percentage of genes on chromosome in that category

Functional categories	<i>Escherichia coli</i> (4.64 Mbp) ^a	<i>Haemophilus influenzae</i> (1.83 Mbp) ^a	<i>Mycoplasma genitalium</i> (0.58 Mbp) ^a
Metabolism	21.0	19.0	14.6
Structural	5.5	4.7	3.6
Transport	10.0	7.0	7.3
Regulation	8.5	6.6	6.0
Translation	4.5	8.0	21.6
Transcription	1.3	1.5	2.6
Replication	2.7	4.9	6.8
Other, known	8.5	5.2	5.8
Unknown	38.1	43.0	32.0

^a Chromosome size. Each organism listed contains only a single circular chromosome.

Genome size vs. ORFs

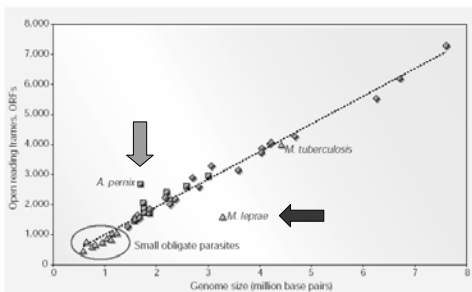


Figure 1 Number of genes (ORFs) plotted against genome size for 44 fully sequenced genomes, including ten Archaea (squares) and 34 Bacteria. Obligate bacterial parasites are denoted by triangles; all other bacteria are shown as diamonds. *Mycobacterium leprae* is a genome 'in decay' that has a large number of pseudogenes. The archaeon *Aeropyrum pernix* is unusual in having an excessive number of duplicated ORFs.

Organism (number of genes)	Glycolysis	Tricarboxylic acid cycle	Amino acid biosynthesis	Purine biosynthesis	Pyrimidine biosynthesis	Ancestral stock
<i>Mycoplasma genitalium</i> (470)	+	-	-	-	-	<i>Bacillus-Clostridium</i>
<i>Buchnera</i> species (588)	+	-	+	+	+	Gamma- proteobacteria
<i>Rickettsia prowazekii</i> (834)	-	+	-	-	-	Alpha- proteobacteria
<i>Chlamydia trachomatis</i> (894)	+	-	+	-	-	Main line
<i>Treponema pallidum</i> (1,041)	+	-	-	-	-	Main line
<i>Mycobacterium leprae</i> (1,604)	Partial	In decay	+	+	+	<i>Bacillus-Clostridium</i>

Figure 2 Many routes to intracellular adaptation. The differing presence (+) or absence (-) of certain metabolic pathways in the streamlined genomes of parasitic bacteria shows how variable the process may be.

Global Transposon Mutagenesis and a Minimal Mycoplasma Genome

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Robin T. Cline,¹ Owen White,¹ Claire M. Fraser,¹
Hamilton O. Smith,^{1,‡} J. Craig Venter^{1,§}

Mycoplasma genitalium with 517 genes has the smallest gene complement of any independently replicating cell so far identified. Global transposon mutagenesis was used to identify nonessential genes in an effort to learn whether the naturally occurring gene complement is a true minimal genome under laboratory growth conditions. The positions of 2209 transposon insertions in the completely sequenced genomes of *M. genitalium* and its close relative *M. pneumoniae* were determined by sequencing across the junction of the transposon and the genomic DNA. These junctions defined 1354 distinct sites of insertion that were not lethal. The analysis suggests that 265 to 350 of the 480 protein-coding genes of *M. genitalium* are essential under laboratory growth conditions, including about 100 genes of unknown function.

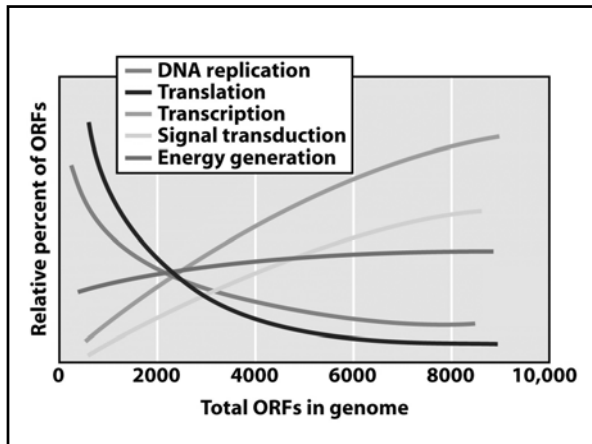
265 to 350 genes are the minimum necessary genome

The complete genome of the hyperthermophilic bacterium *Aquifex aeolicus*

Gerard Deckert¹, Patrick V. Warren¹, Terry Gaasterland¹, William G. Young¹, Anna L. Lenox¹, David E. Graham¹, Ross Overbeek¹, Marjory A. Smead¹, Martin Keller¹, Monette Aujay¹, Robert Hubert¹, Robert A. Feldman¹, Jay M. Short¹, Gary J. Olsen¹ & Ronald V. Swanson¹

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Aquifex aeolicus was one of the earliest diverging, and is one of the most thermophilic, bacteria known. It can grow on hydrogen, oxygen, carbon dioxide, and mineral salts. The complex metabolic machinery needed for *A. aeolicus* to function as a chemolithoautotroph (an organism which uses an inorganic carbon source for biosynthesis and an inorganic chemical energy source) is encoded within a genome that is only one-third the size of the *E. coli* genome. Metabolic flexibility seems to be reduced as a result of the limited genome size. The use of oxygen (albeit at very low concentrations) as an electron acceptor is allowed by the presence of a complex respiratory apparatus. Although this organism grows at 95 °C, the extreme thermal limit of the Bacteria, only a few specific indications of thermophily are apparent from the genome. Here we describe the complete genome sequence of 1,551,335 base pairs of this evolutionarily and physiologically interesting organism.

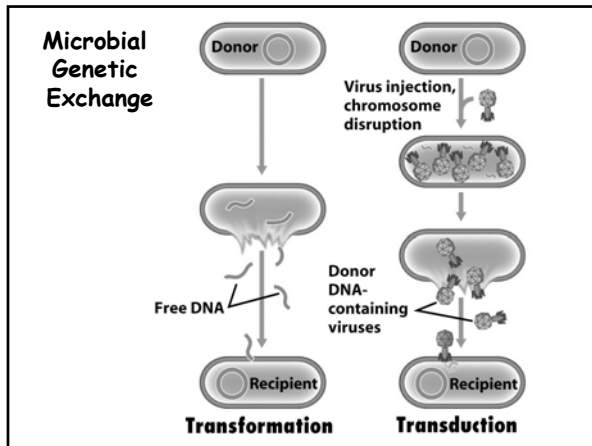


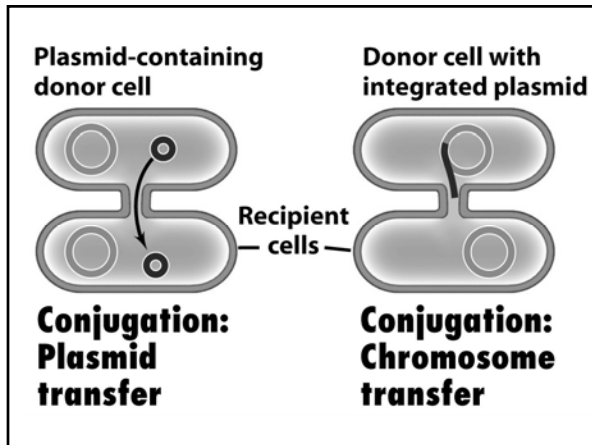
Lessons from full genomes

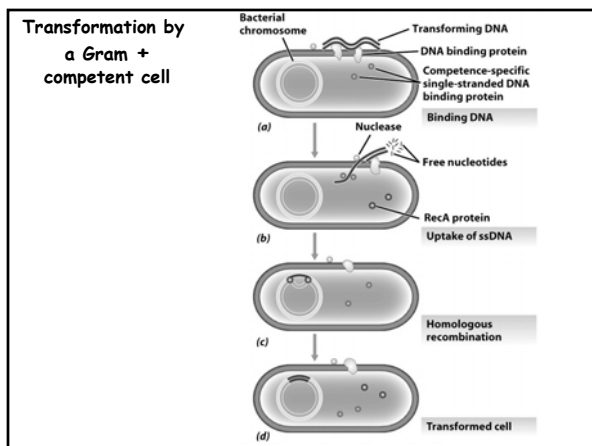
- Size range 600Kb to 12Mb
- Vast number of putative genes with no known function
- Pathogenicity conferred by "Pathogenicity Islands"
44.5Kb in *Bacillus anthracis*
- Symbiotic Island of >600Kb in *Sinorhizobium loti*
including genes for nodulation and N-fixation
- Adaptive gene losses in parasitic bacteria
Rickettsia and *Chlamidia* are ATP thieves using
the same "alien" ADP/ATP translocase
- Relative proportions of functional genes

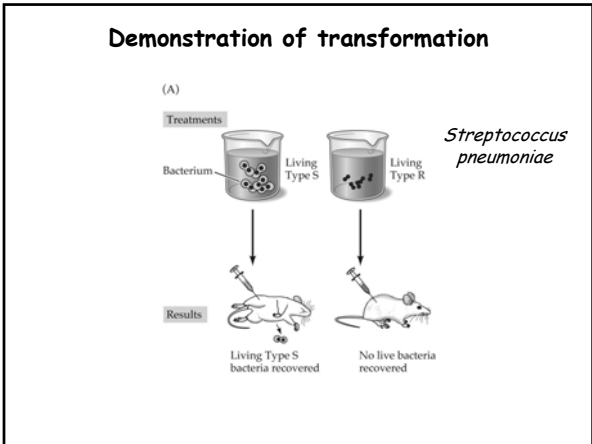
Microbial Genetic Exchange and Plasmids

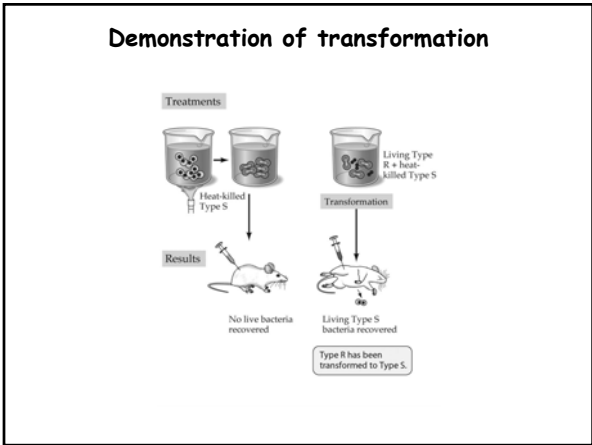
- Microbial Genetic Exchange is unidirectional!
 - Transformation
 - Transduction
 - Conjugation
- Each requires Homologous Recombination
- Types of plasmids

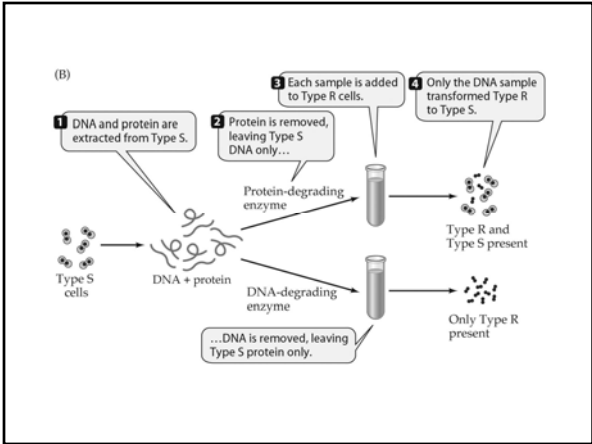




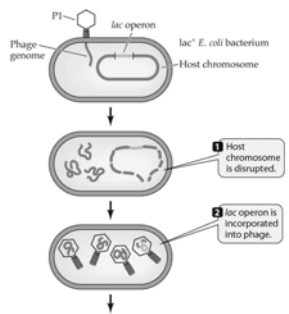




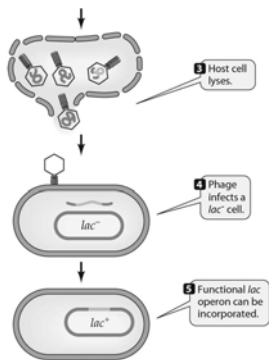




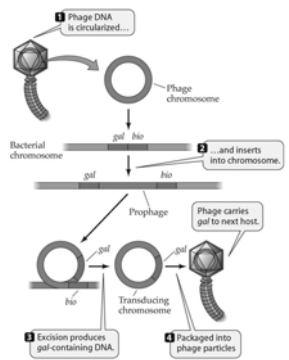
Generalized transduction

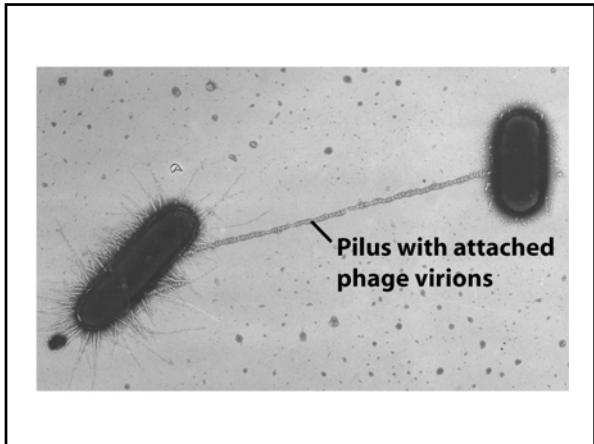


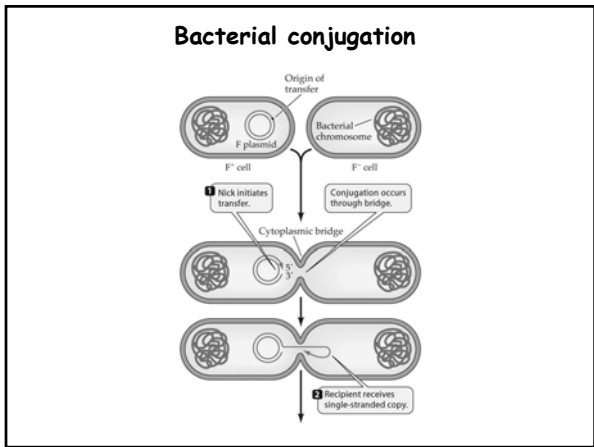
Generalized transduction (cont.)

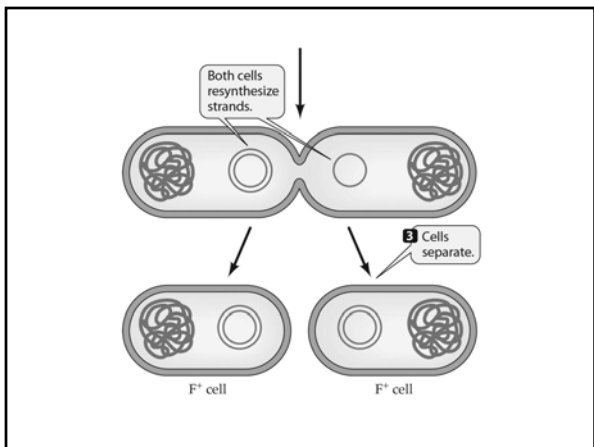


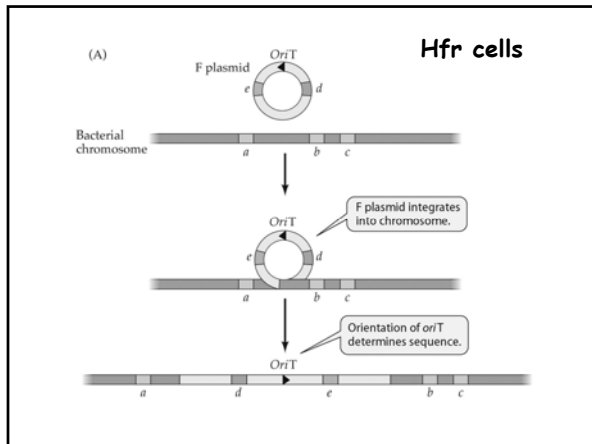
Specialized transduction

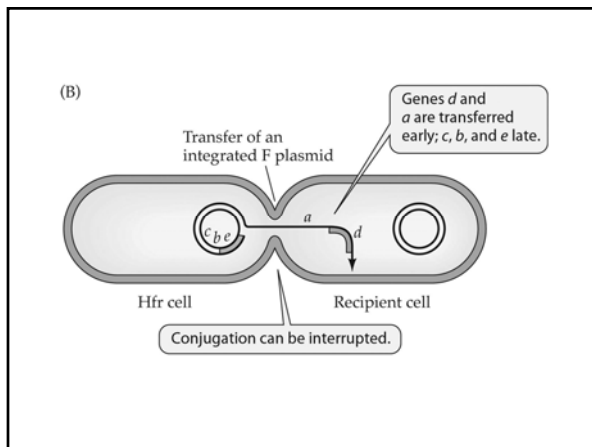


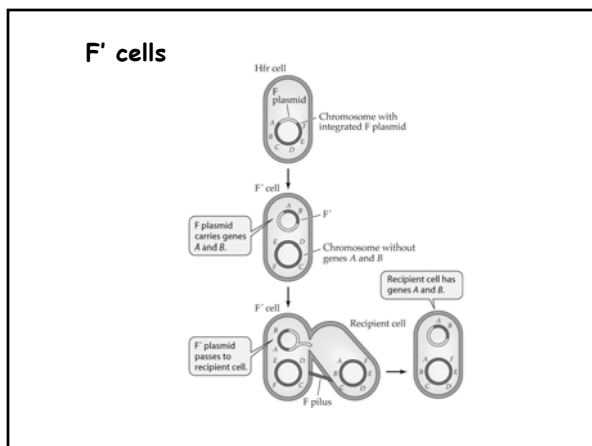


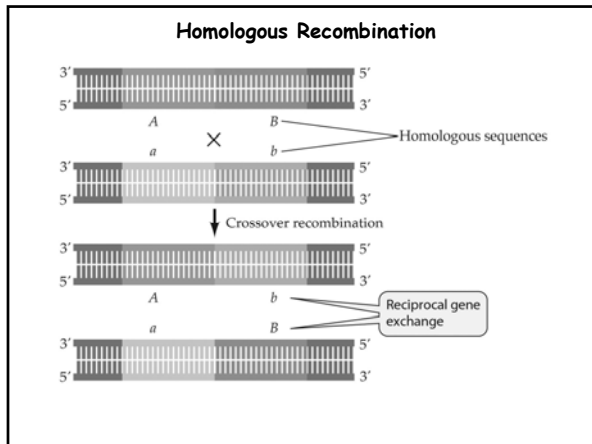


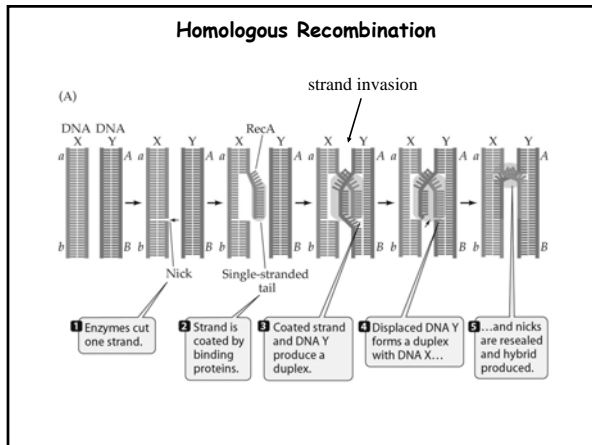


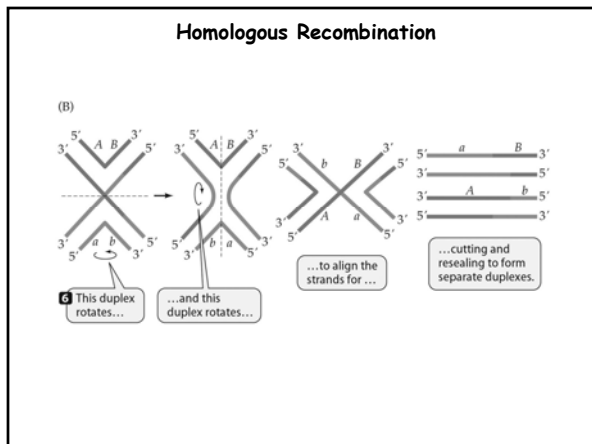




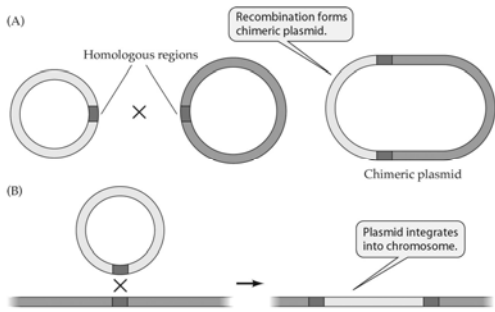




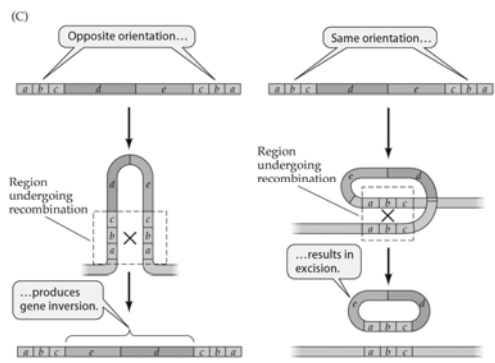




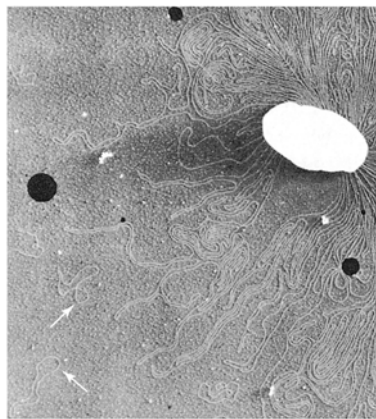
Types of homologous recombination in bacteria

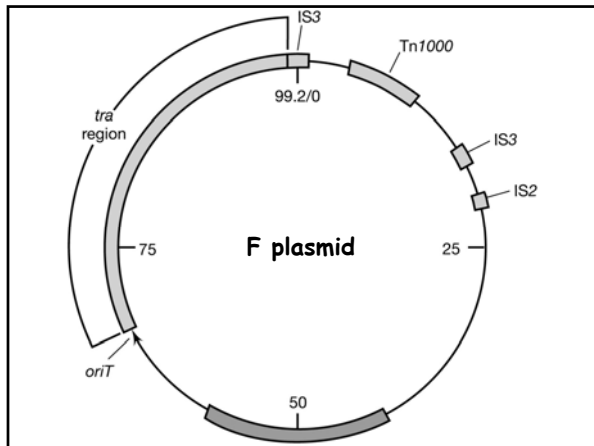


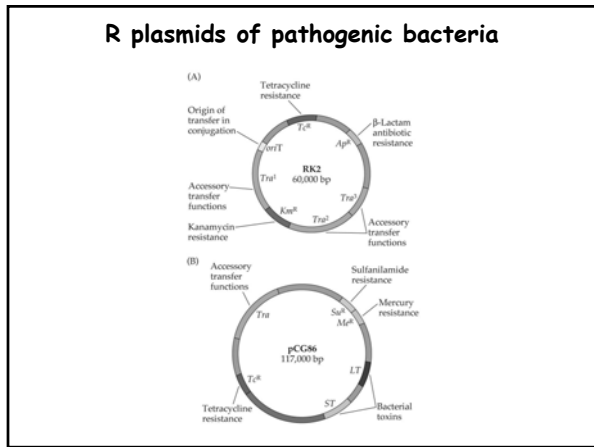
Types of homologous recombination in bacteria



Plasmids







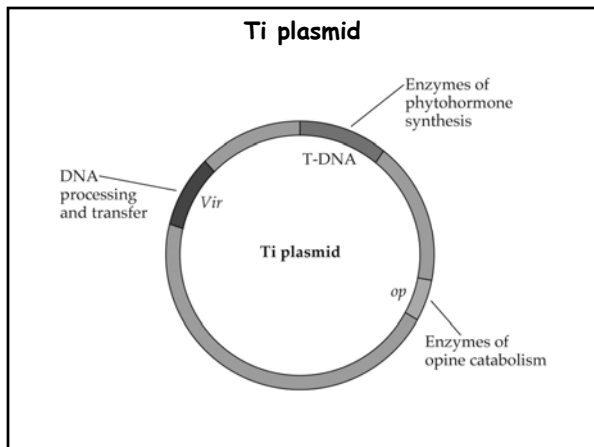


Table 10.3 Some phenotypes conferred by plasmids in prokaryotes

Phenotype class ^a	Organisms ^b
Antibiotic production	Streptomycetes
Conjugation	<i>Escherichia</i> , <i>Pseudomonas</i> , <i>Rhizobium</i> , <i>Staphylococcus</i> , <i>Streptococcus</i> , <i>Sulfolobus</i> , <i>Vibrio</i>
Physiological functions	
Degradation of octane, camphor, naphthalene	<i>Pseudomonas</i>
Degradation of herbicides	<i>Alcaligenes</i>
Formation of acetone and butanol (OFD Section 12.20)	<i>Clostridium</i>
Lactose, sucrose or urea utilization and nitrogen fixation	Enteric bacteria
Nodulation and symbiotic nitrogen fixation (OFD Section 19.22)	<i>Rhizobium</i>
Pigment production	<i>Erwinia</i> , <i>Staphylococcus</i>
Resistance	
Antibiotic resistance (OFD Section 20.12)	<i>Campylobacter</i> , Enteric bacteria, <i>Neisseria</i> , <i>Staphylococcus</i>
Resistance to cadmium, cobalt, mercury, nickel, and/or zinc (OFD Sections 19.16)	<i>Acidithiobacillus</i> , <i>Alcaligenes</i> , <i>Listeria</i> , <i>Pseudomonas</i> , <i>Staphylococcus</i>
Bacteriocin resistance (and production)	<i>Bacillus</i> , Enteric bacteria, <i>Lactococcus</i> , <i>Propionibacterium</i>
Virulence	
Host cell invasion	<i>Salmonella</i> , <i>Shigella</i> , <i>Yersinia</i>
Conglutinase, hemolysin, enterotoxin (OFD Sections 21.9 and 21.11)	<i>Staphylococcus</i>
Enterotoxin, K antigen (OFD Sections 12.11 and 21.11)	<i>Escherichia</i>
Tumorigenicity in plants (OFD Section 19.21)	<i>Agrobacterium</i>

^a Only a few of the many phenotypes known to be associated with plasmids are given.

^b Only a few well-characterized examples are given. All of the organisms given in the list are bacteria except for *Sulfolobus*, which is a member of the *Archaea*.
