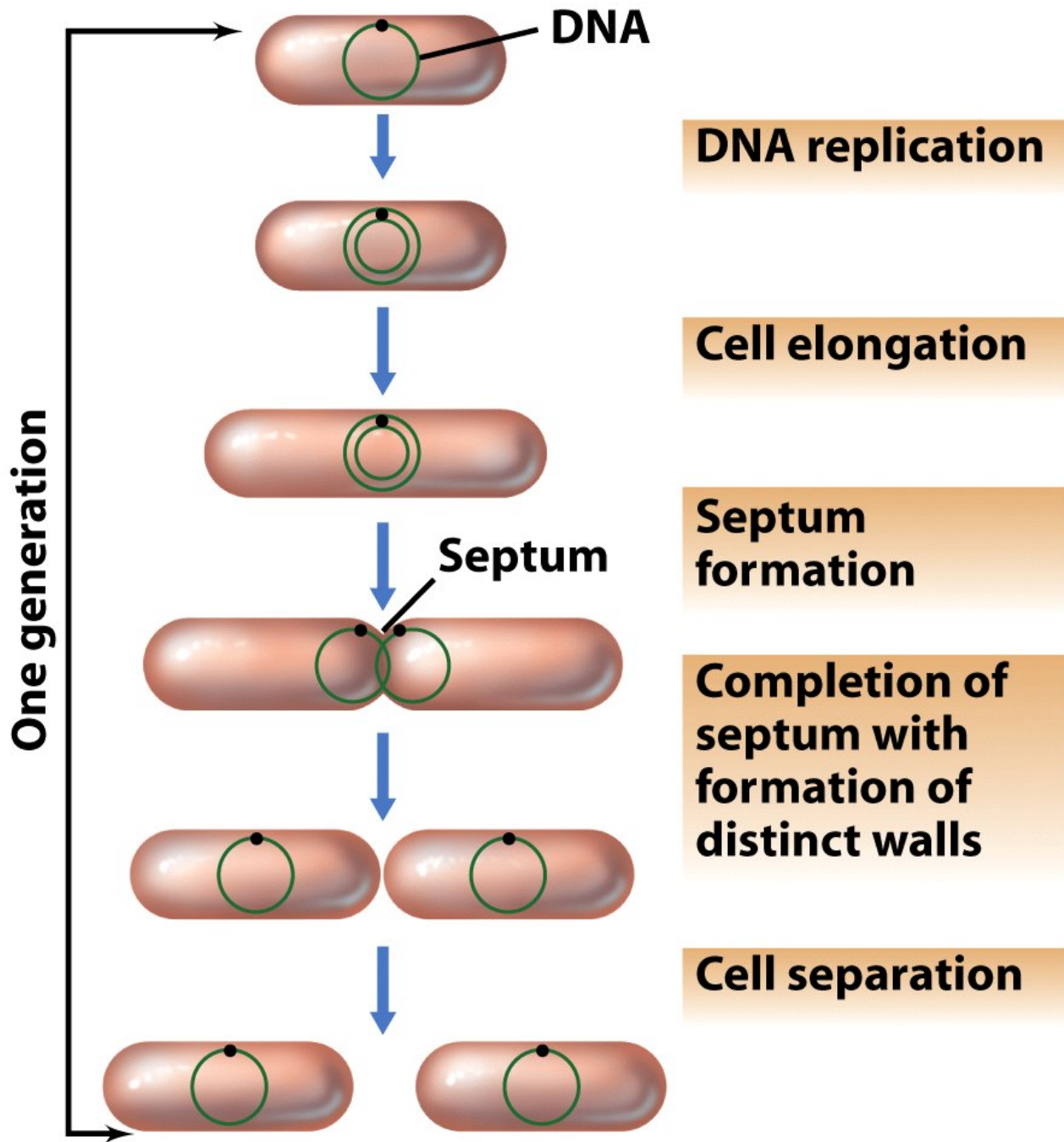


The Process of Growth

- Metabolism required for growth, both anabolic and catabolic. ~2000 reactions!
- Usual Definition: **Increase in cell numbers**
Other definitions possible - spores, UMC's, respiration, viable but nonculturable, morphology changes (life cycle)
- Divide via Binary Fission: 3 mechanisms involved!
Cell Elongation - cell wall
DNA Replication - rate limiting step
Cell Division - septum formation



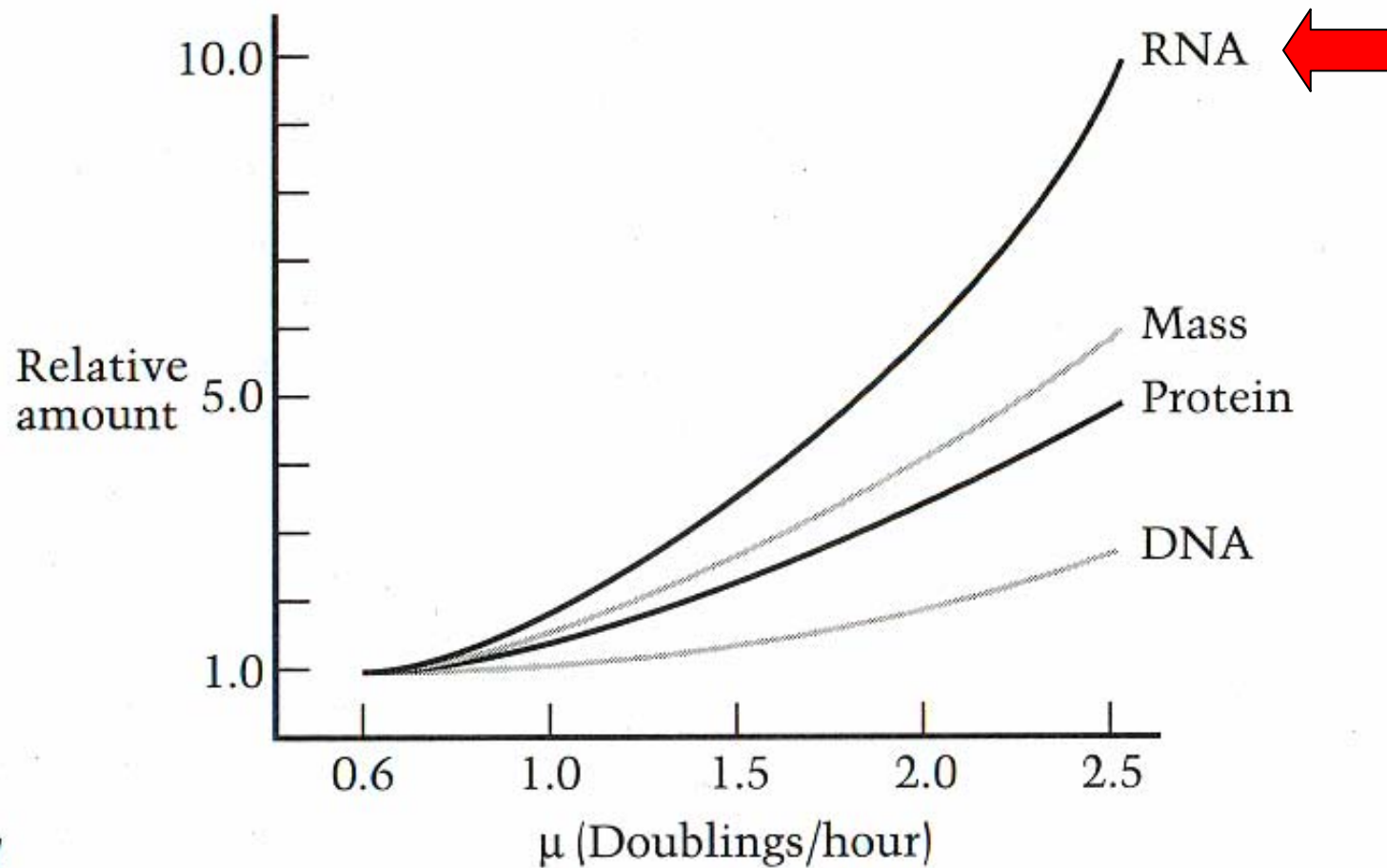


Figure 1

Effect of nutrition-imposed growth rate on the composition of *Escherichia coli* B/r. All values are expressed in amounts per cell normalized to values at $\mu = 0.6$ (mass = 1.48×10^{-13} g; protein = 1.00×10^{-13} g; RNA = 2.0×10^{-14} g; DNA = 6.3×10^{-15} g). (Plotted from data in Bremer and Dennis, 1987.)

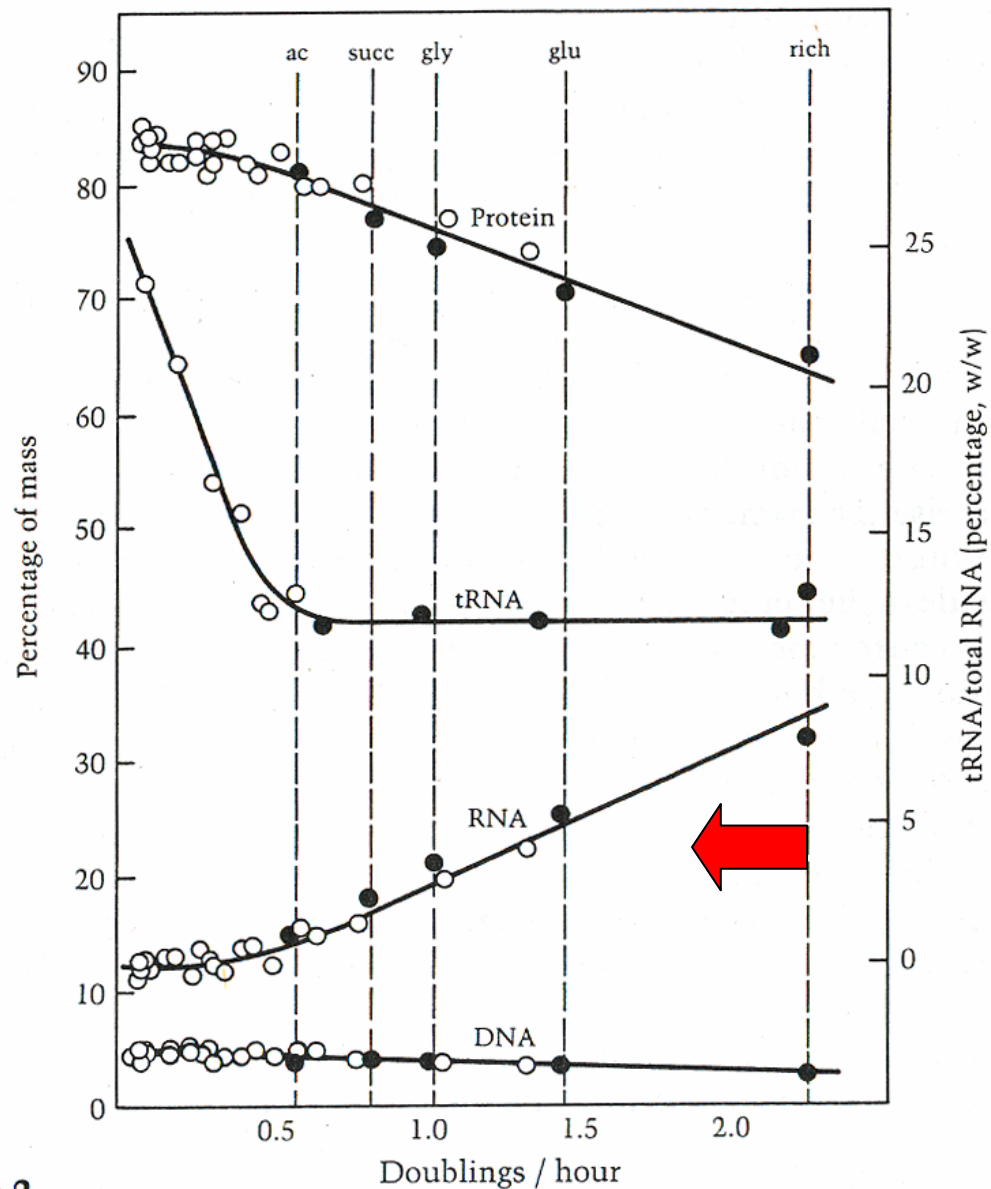
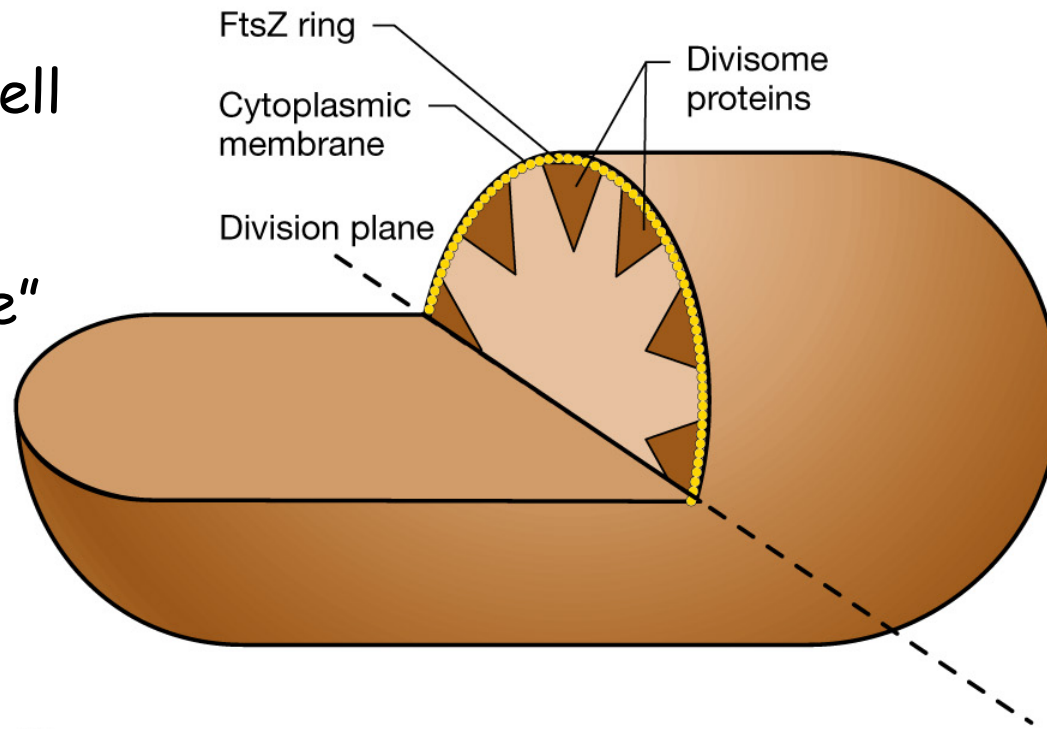


Figure 2

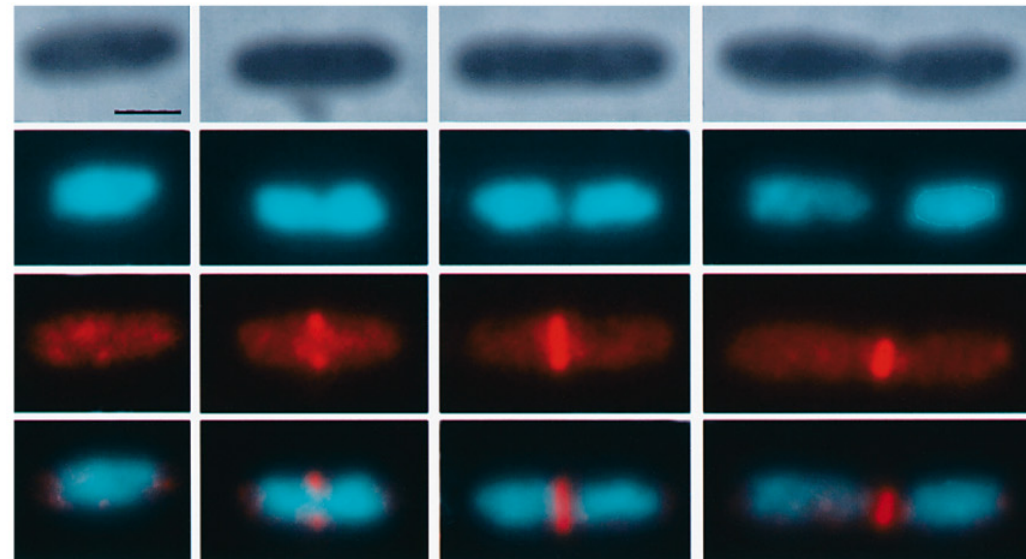
Effect of growth rate on the cellular proportions of protein, RNA, and DNA. Filled circles refer to results from cultures undergoing balanced growth in batch culture in various media; open circles are from cultures growing in a glucose-limited chemostat. (From Jacobsen, 1974.)

FtsZ ring & cell division

The "divisome"



(a)



T. den Blaauwen & Nanne Nanninga, Univ. of Amsterdam

phase

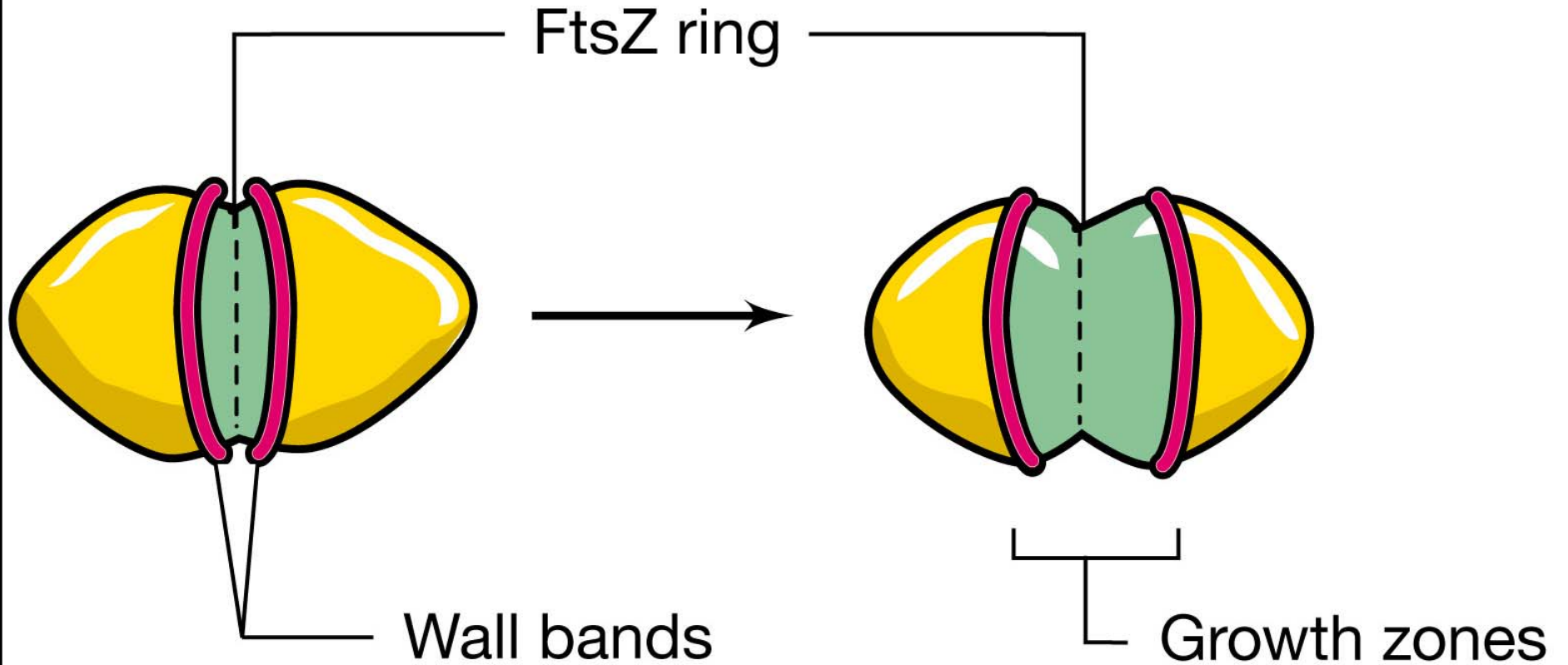
DNA stain

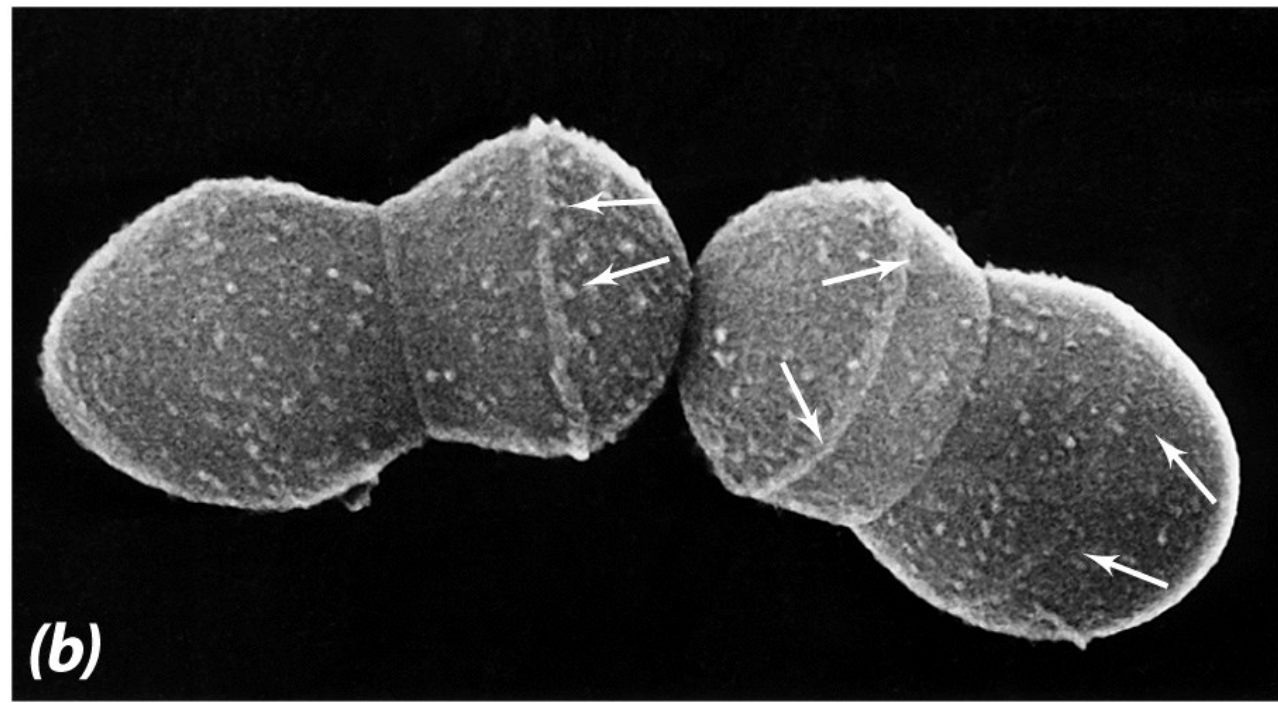
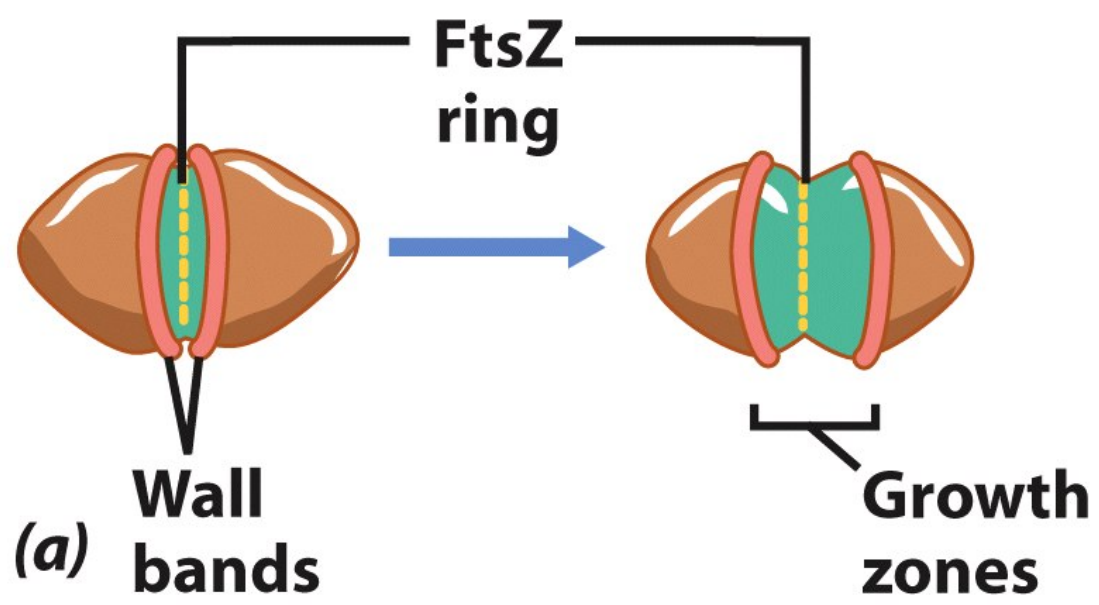
FtsZ stain

DNA & FtsZ

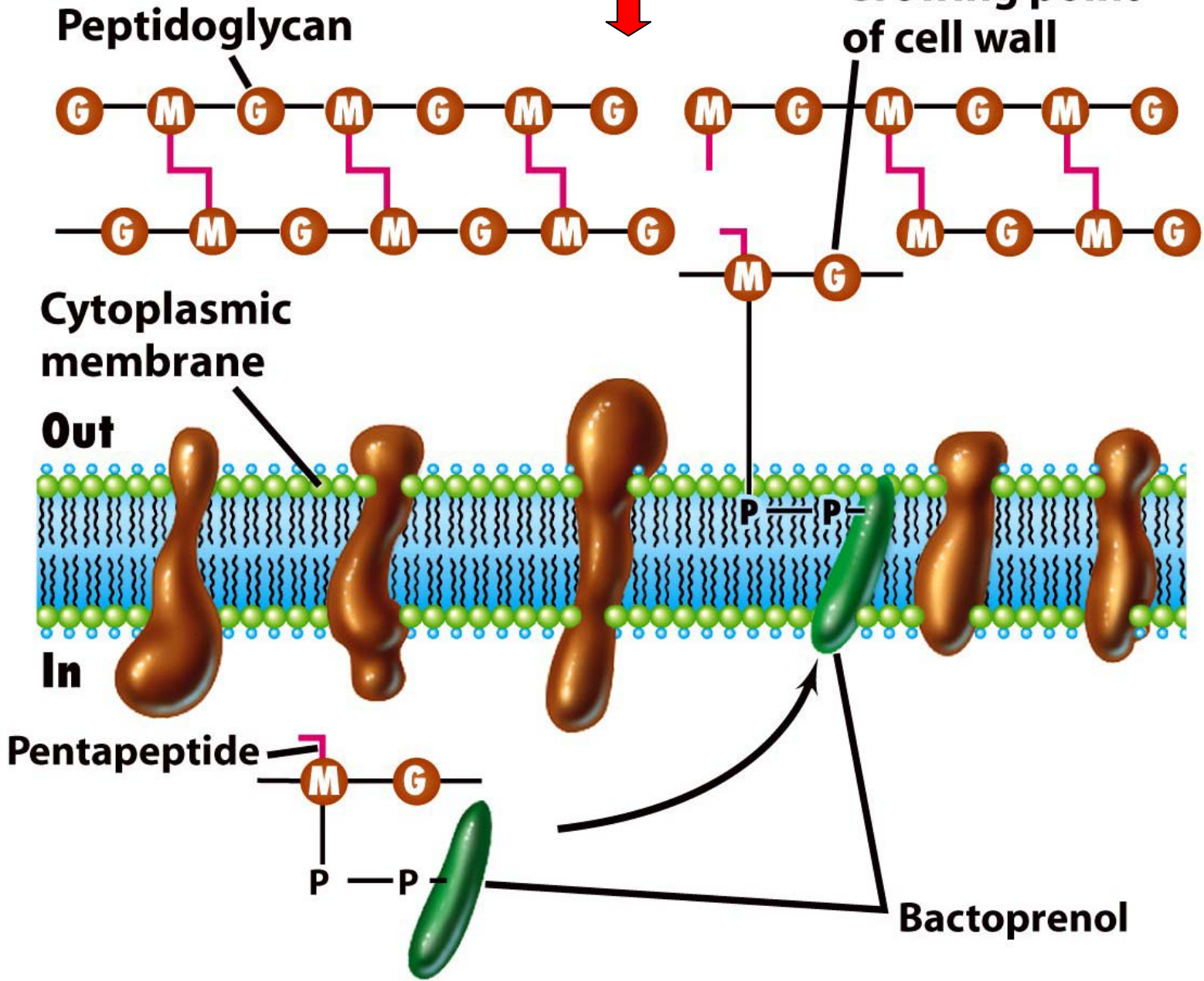
(b)

FtsZ similar to Tubulin
MreB similar to Actin

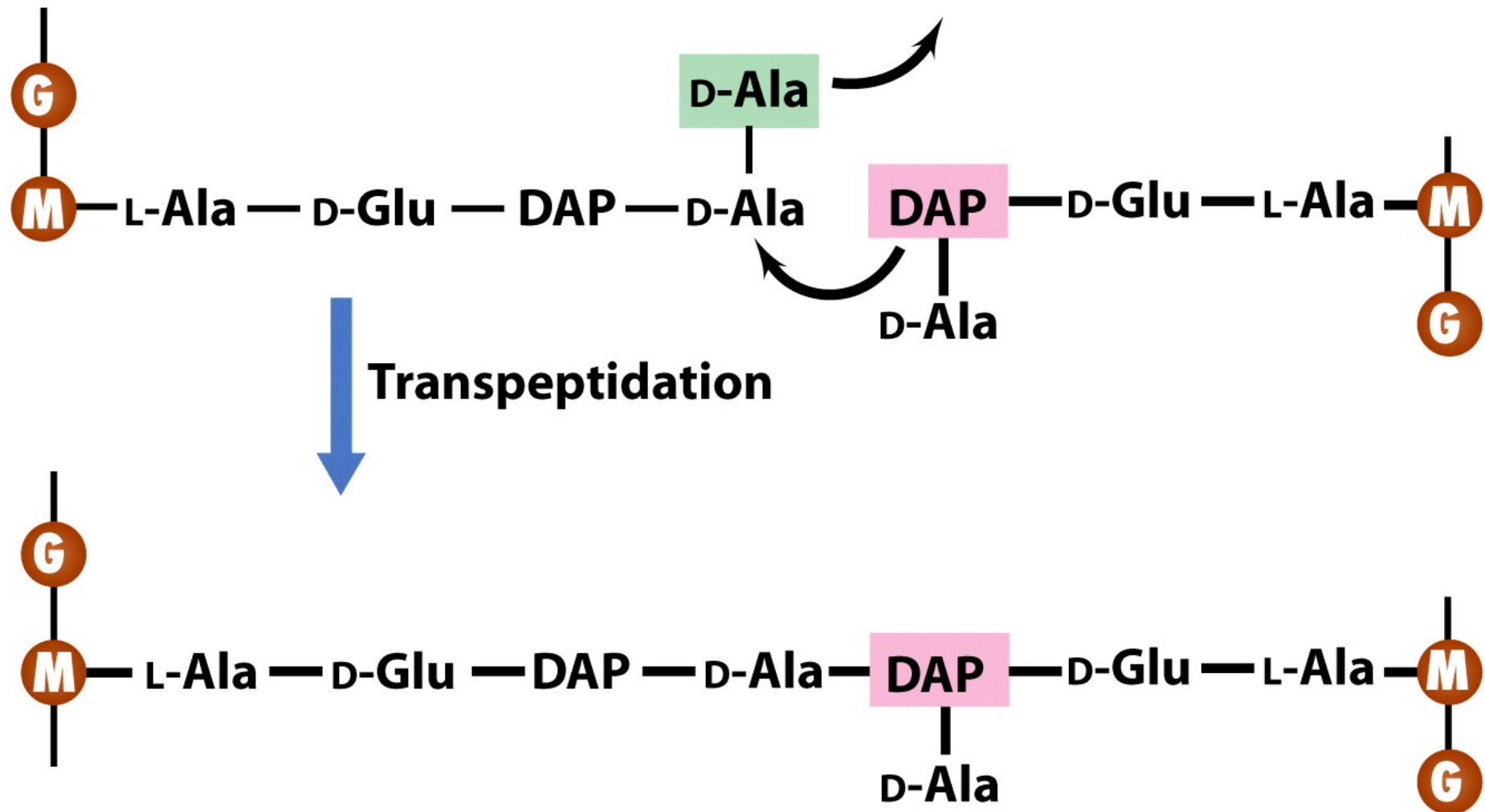




Autolysins



Penicillin blocks this reaction



$\tau_d = 100 \text{ min}$

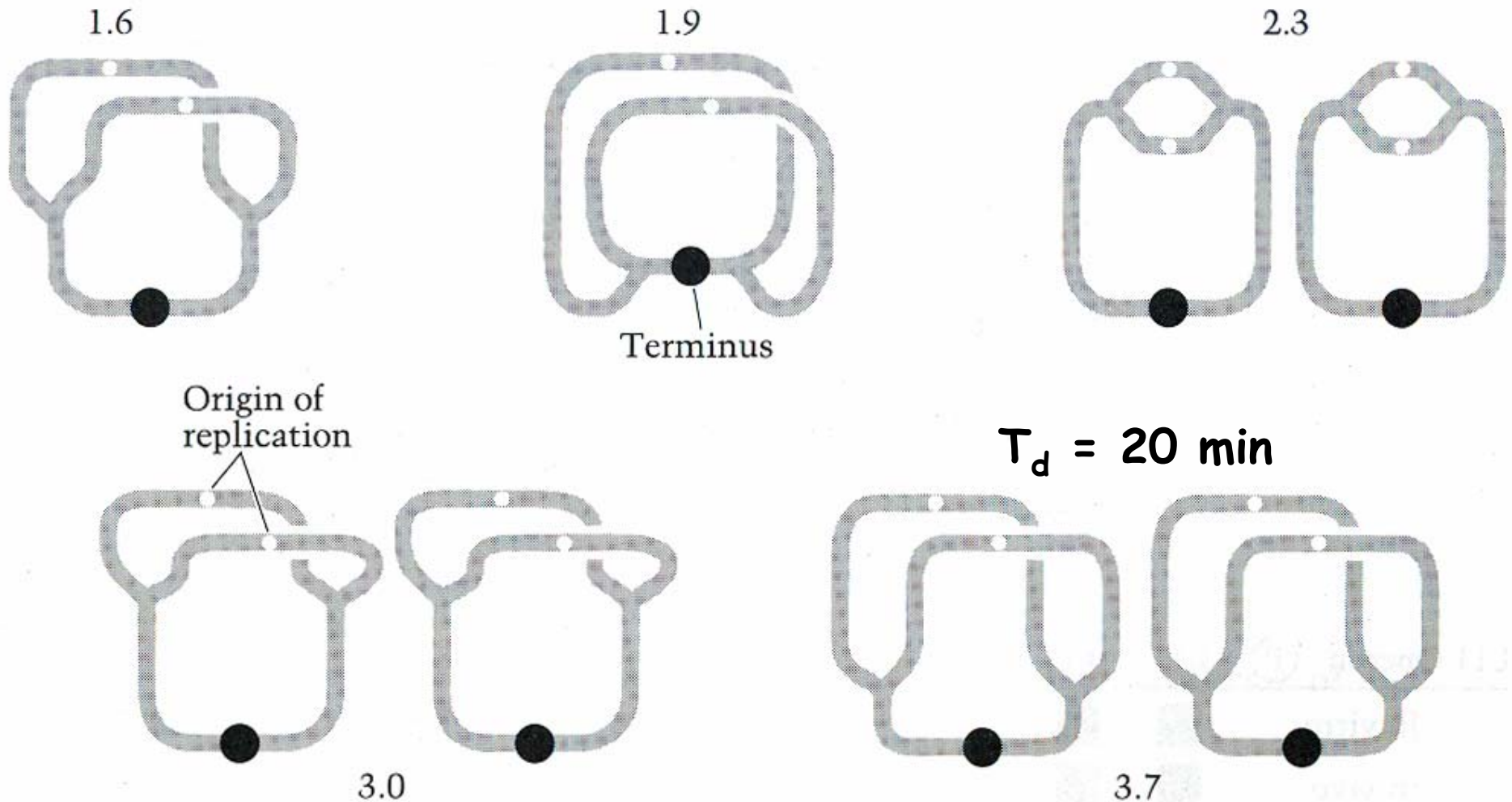
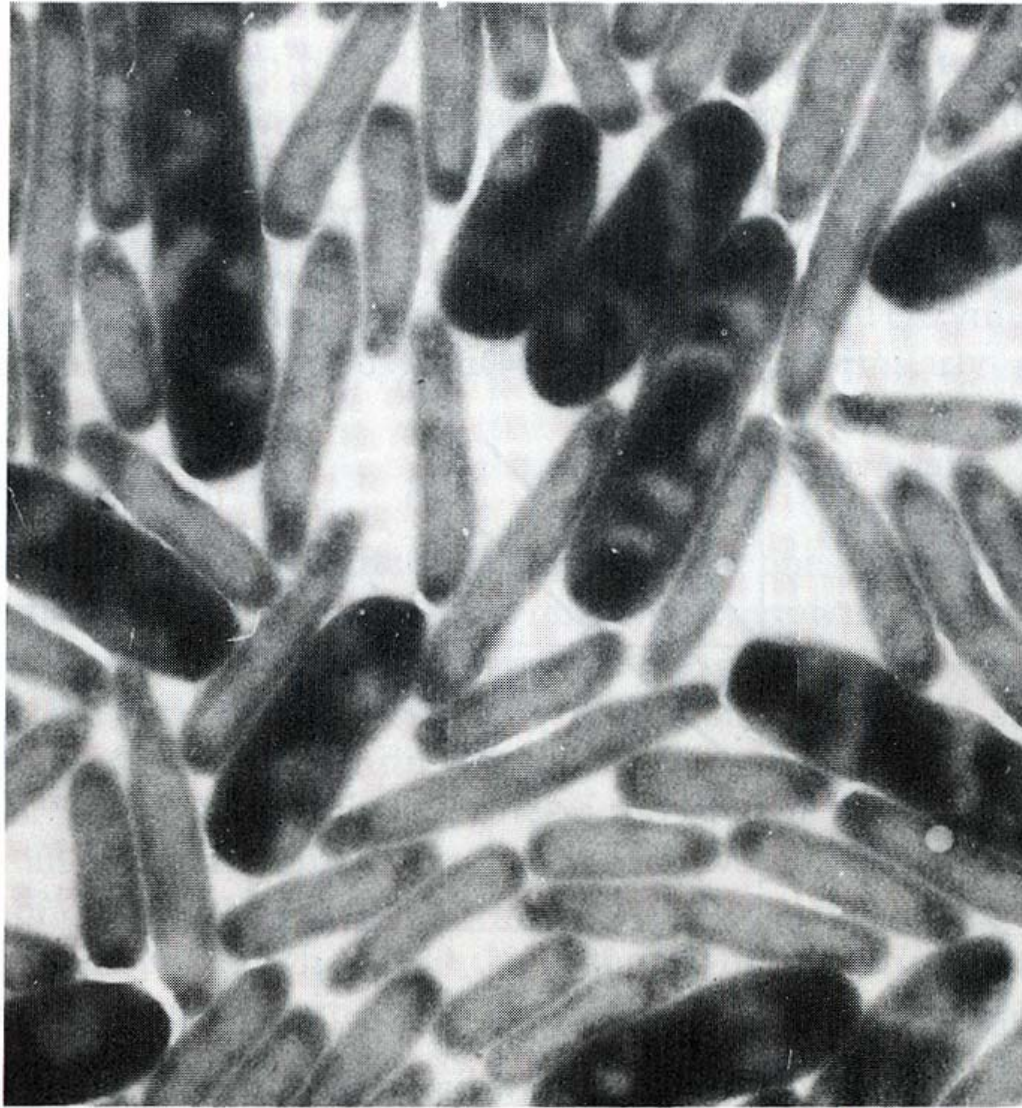


Figure 10

Chromosome structure and equivalent DNA content of the average cell in culture of *E. coli* B/r growing at various rates. The numbers represent genome equivalents. (From Bremer and Dennis, 1987.)



1 μm

Figure 14

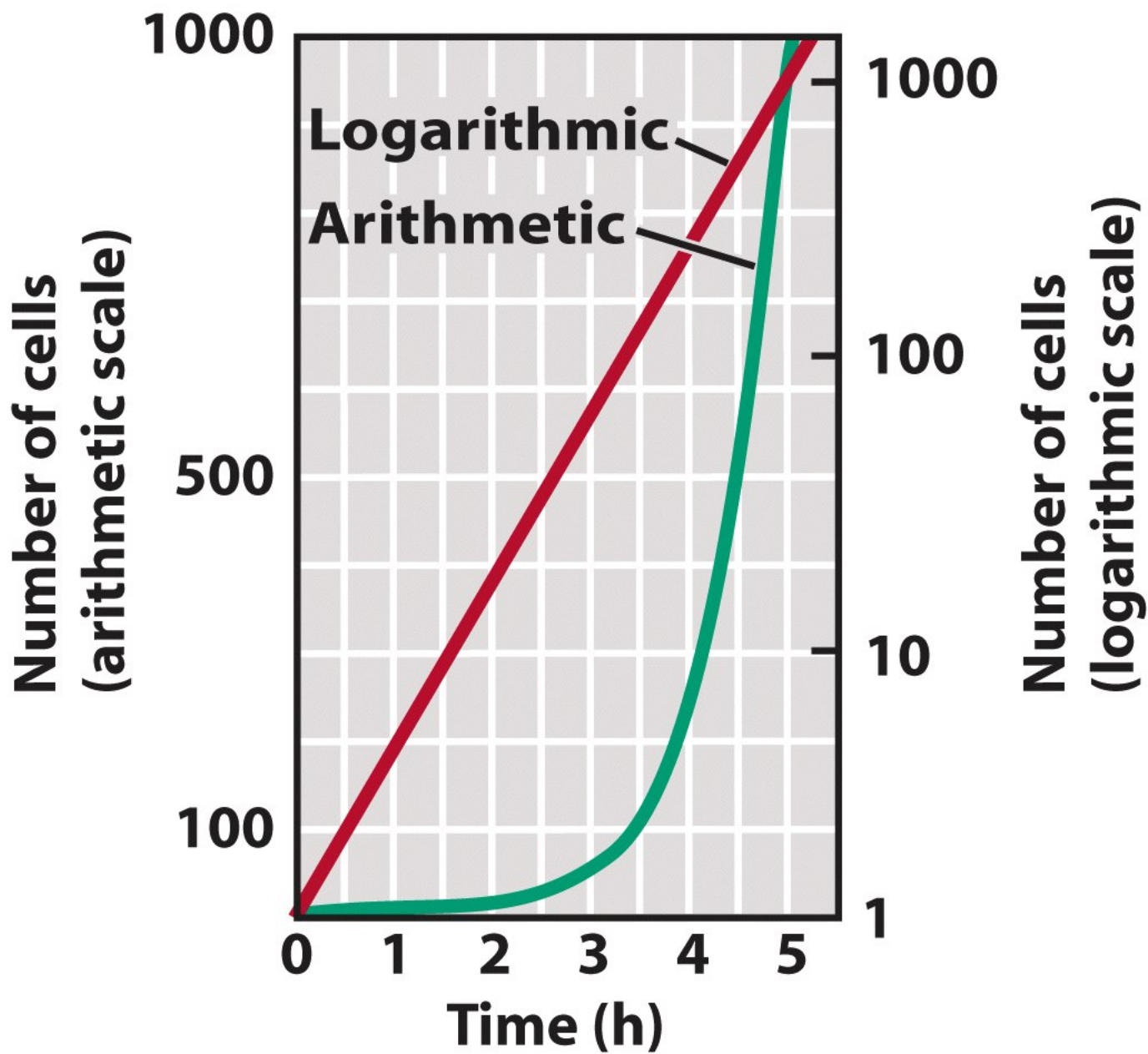
Electron micrograph of a mixture of cells of *E. coli* B/r grown at different rates. The large cells grew with a doubling time of 22 minutes, the small ones with a doubling time of 72 minutes. (From Nanninga and Woldringh, 1985.)

The Process of Growth

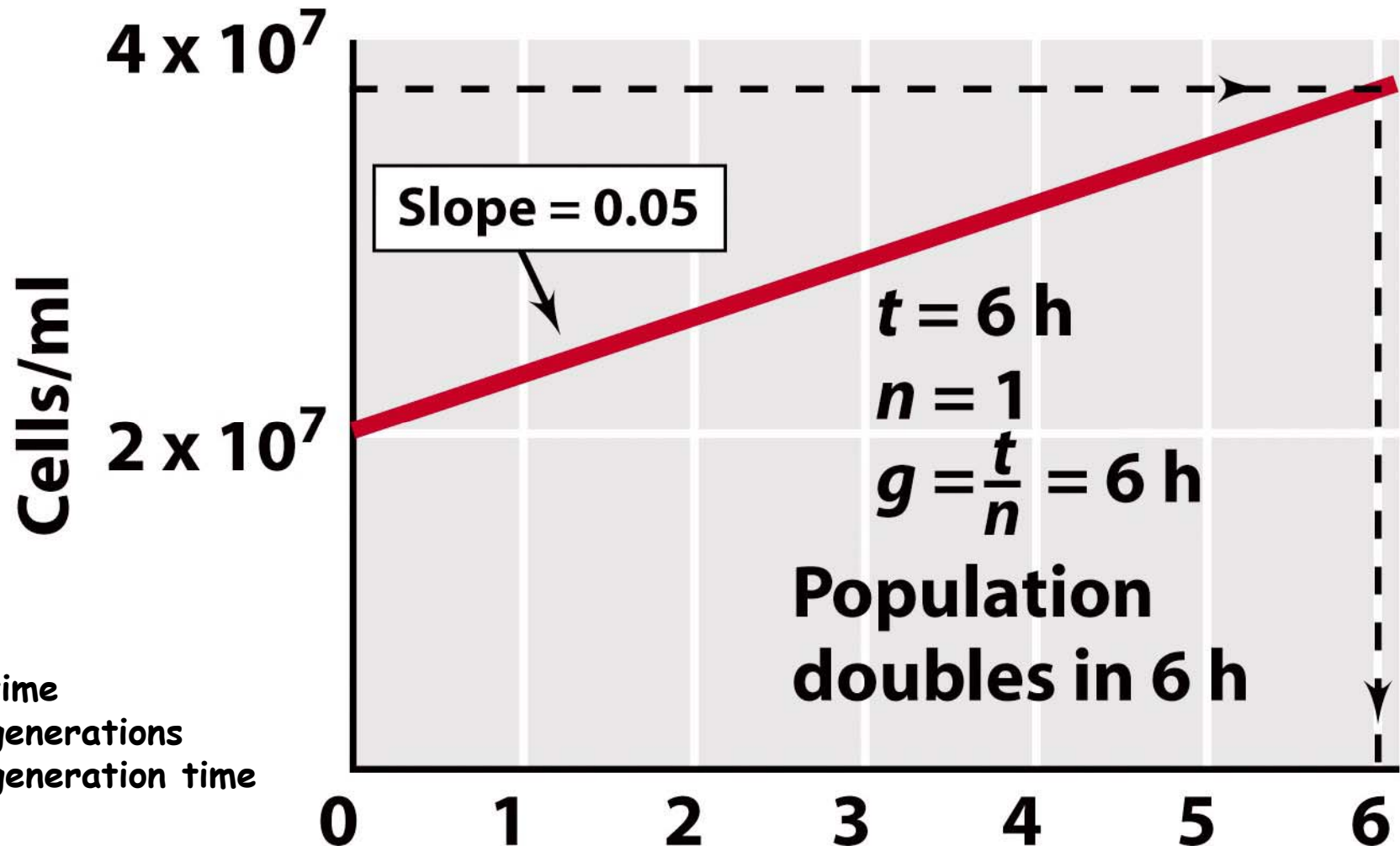
- Growth Rate: Time it takes to reproduce
 $t_{\frac{1}{2}} = \ln 2 / \mu = 0.693 / \mu = g$
- Phases of Growth in Batch culture
Lag, Log, Stationary, Death
- Measurement of Growth
Total cell counts
Viable cell counts
Turbidity

The growth rate of a microbial culture

Time (h)	Total number of cells	Time (h)	Total number of cells
0	1	4	256 (2^8)
0.5	2	4.5	512 (2^9)
1	4	5	1,024 (2^{10})
1.5	8	5.5	2,048 (2^{11})
2	16	6	4,096 (2^{12})
2.5	32	.	.
3	64	.	.
3.5	128	10	1,048,576 (2^{19})



How to estimate the generation times of an exponential microbial culture using semi-log plots.



t = time
n = generations
g = generation time

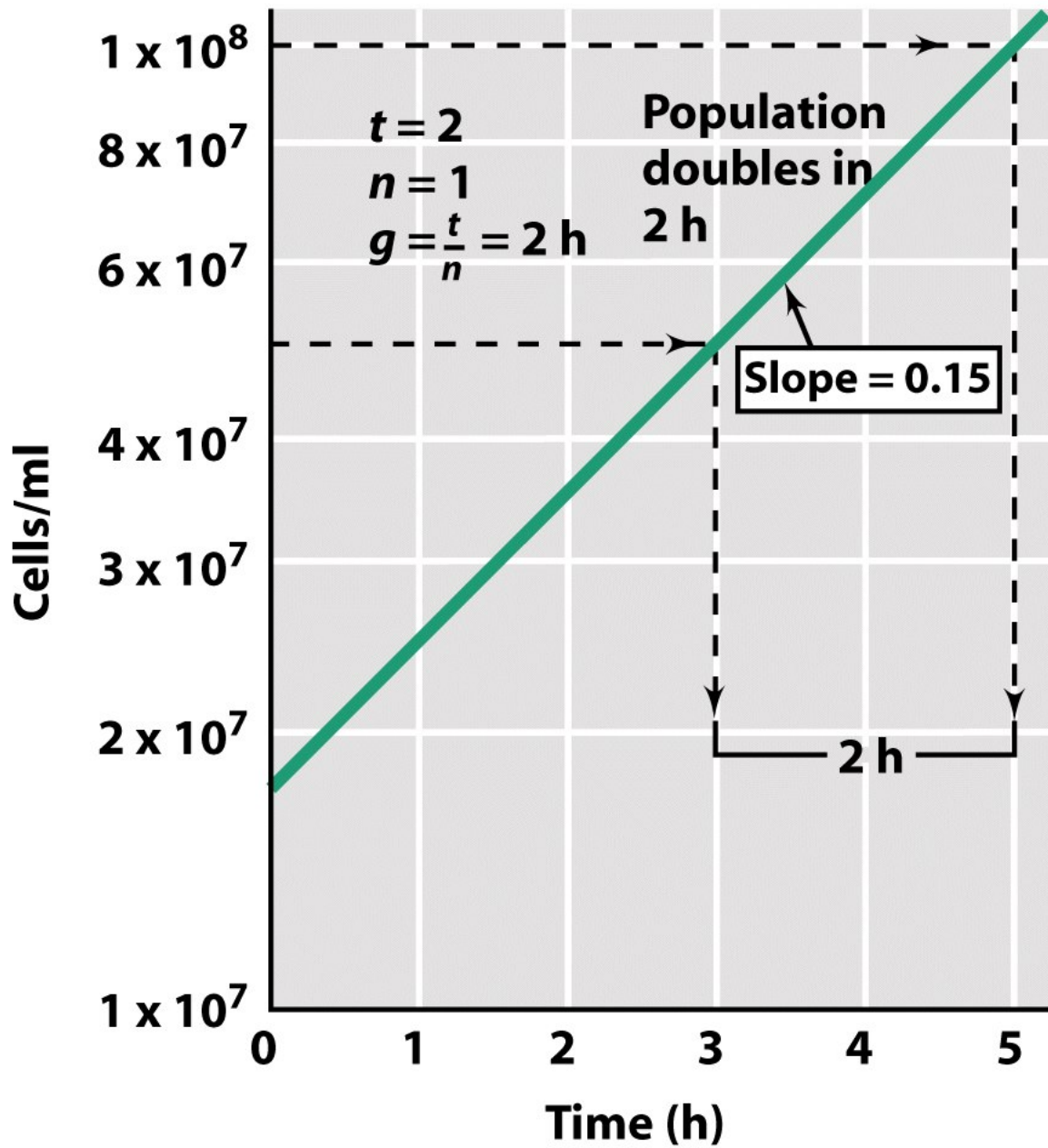
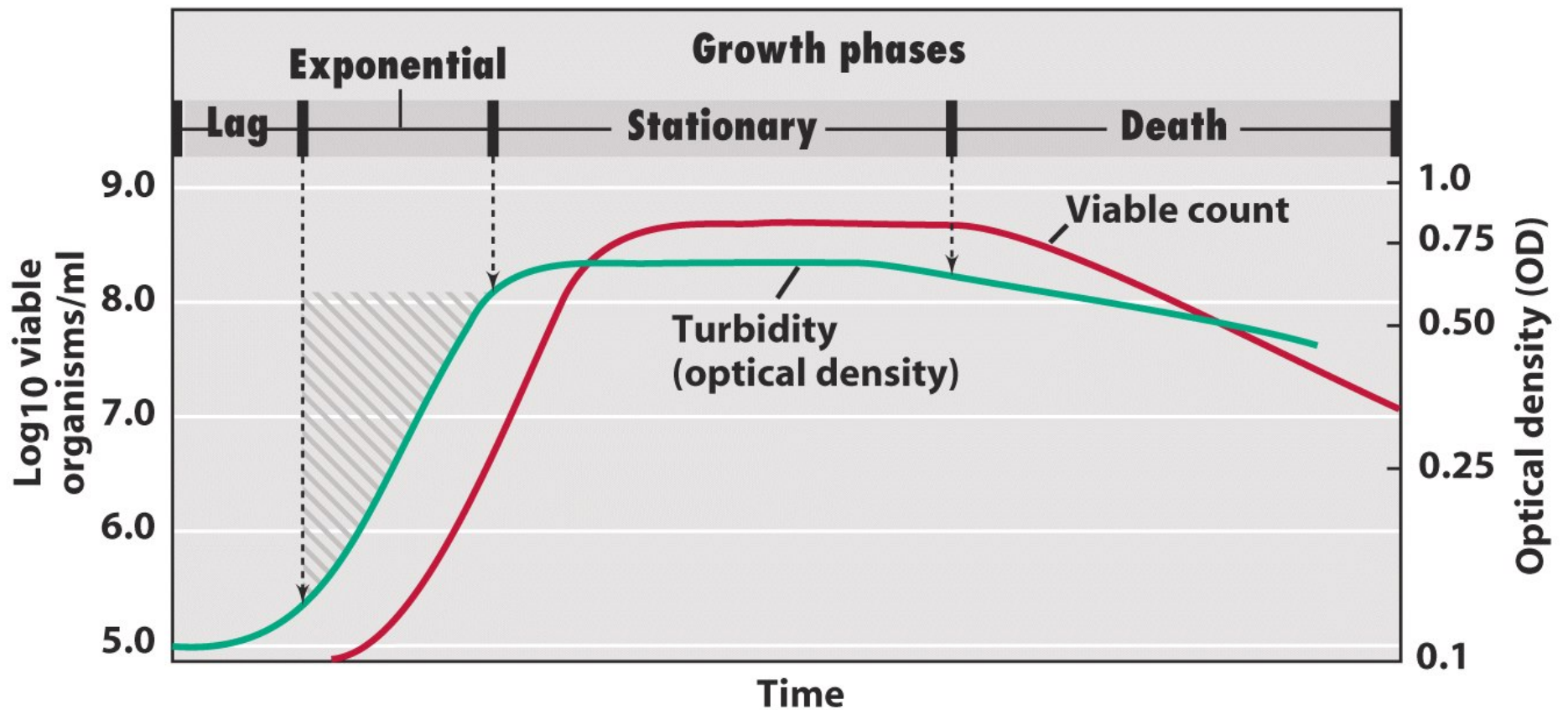


Table 6.1**Approximate generation times for several organisms growing in media optimal for growth**

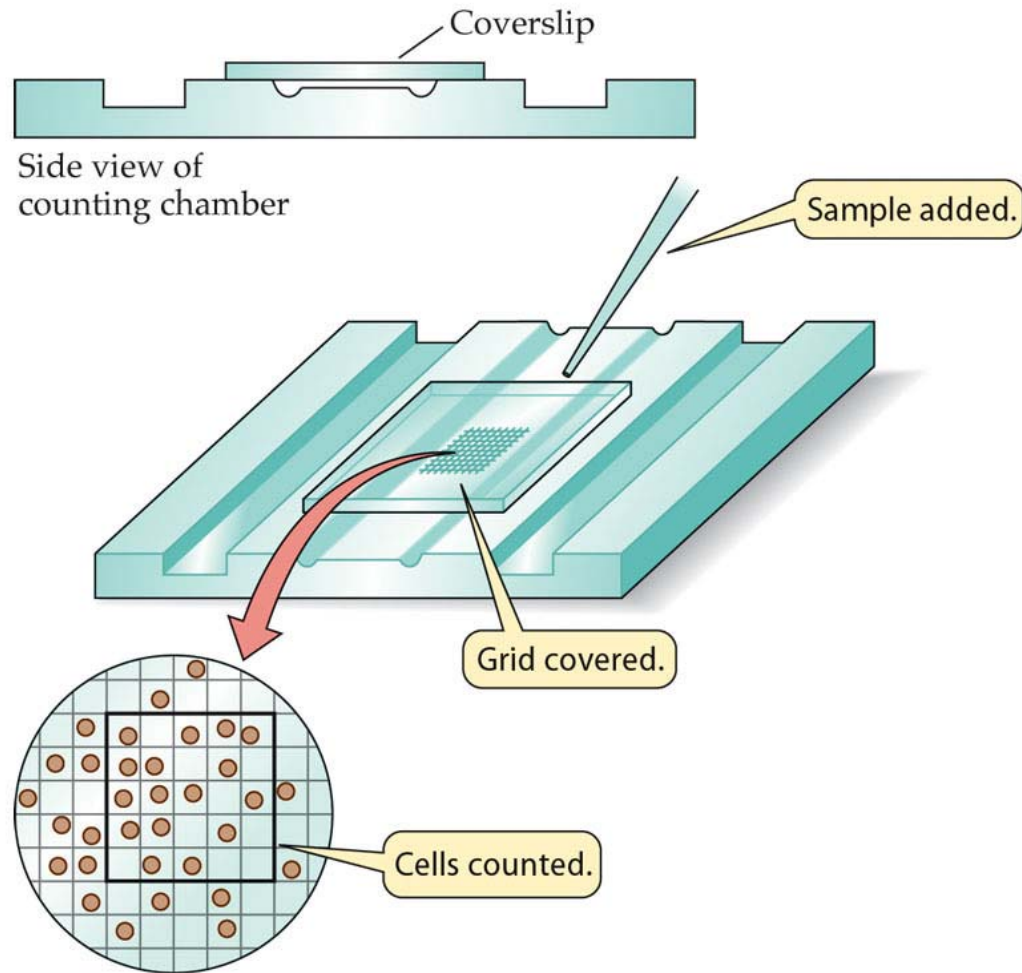
Species	Generation Time
<i>Escherichia coli</i>	20 min
<i>Bacillus subtilis</i>	28 min
<i>Staphylococcus aureus</i>	30 min
<i>Pseudomonas aeruginosa</i>	35 min
<i>Thermus aquaticus</i>	50 min
<i>Thermoproteus tenax</i>	1 hr 40 min
<i>Rhodobacter sphaeroides</i>	2 hr 20 min
<i>Sulfolobus acidocaldarius</i>	4 hr
<i>Thermoleophilum album</i>	6 hr
<i>Thermofilum pendens</i>	10 hr
<i>Mycobacterium tuberculosis</i>	13 hr 20 min

Cryptic Growth ↓

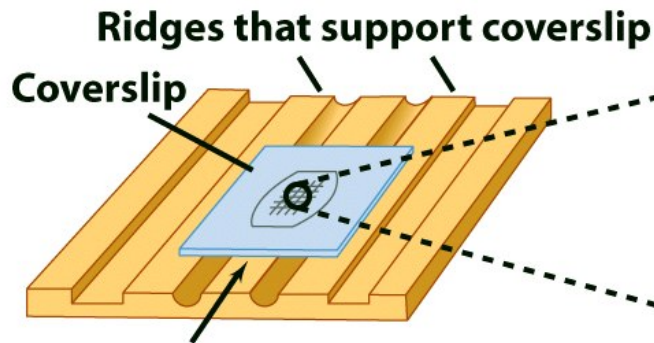


Typical growth curve for a bacterial population

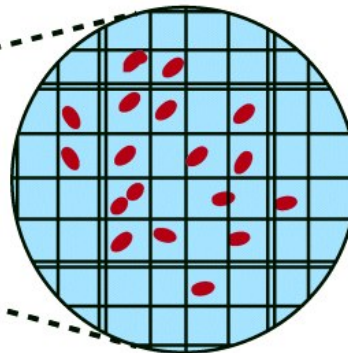
Total Cell counts using the Petroff-Hausser Counter



Total Cell counts using the Petroff-Hausser Counter

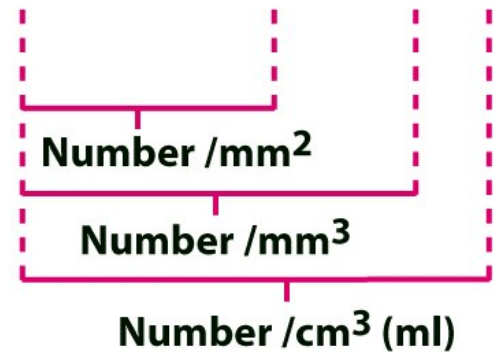


Sample added here; care must be taken not to allow overflow; space between coverslip and slide is 0.02 mm ($\frac{1}{50}$ mm). Whole grid has 25 large squares, a total area of 1 mm² and a total volume of 0.02 mm³.



Microscopic observation; all cells are counted in large square: 12 cells (in practice, several squares are counted and the numbers averaged.)

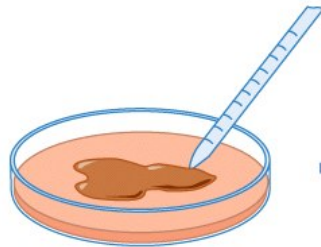
To calculate number per milliliter of sample:
12 cells x 25 large squares
x 50 x 10³ = 1.5 x 10⁷



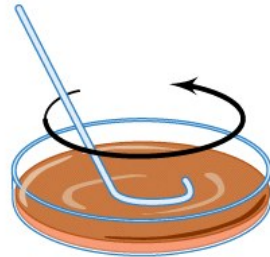
Viabile cell count methods

30-300 on standard
Petri Dish

Spread-plate method

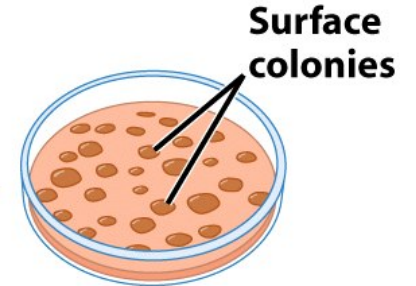
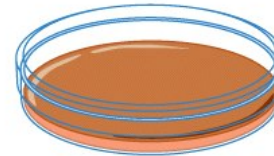


Sample is pipetted
onto surface of agar
plate (0.1 ml or less)



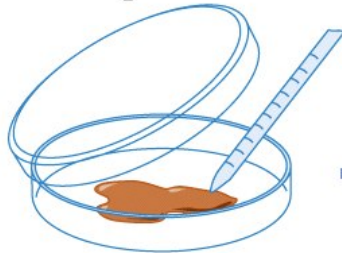
Sample is spread evenly over
surface of agar using sterile
glass spreader

Incubation

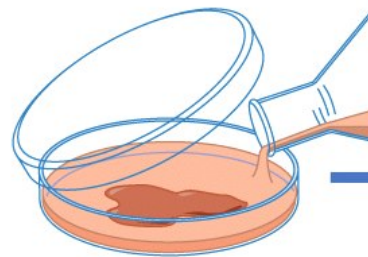


Typical spread-plate
results

Pour-plate method

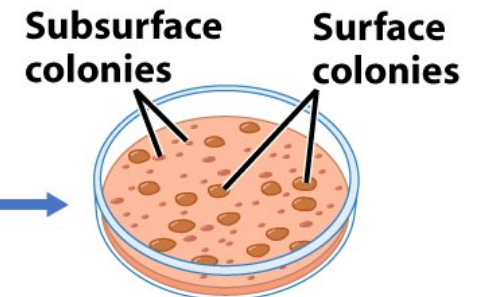
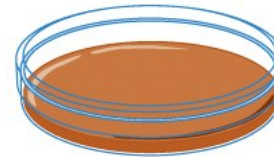


Sample is pipetted
into sterile plate



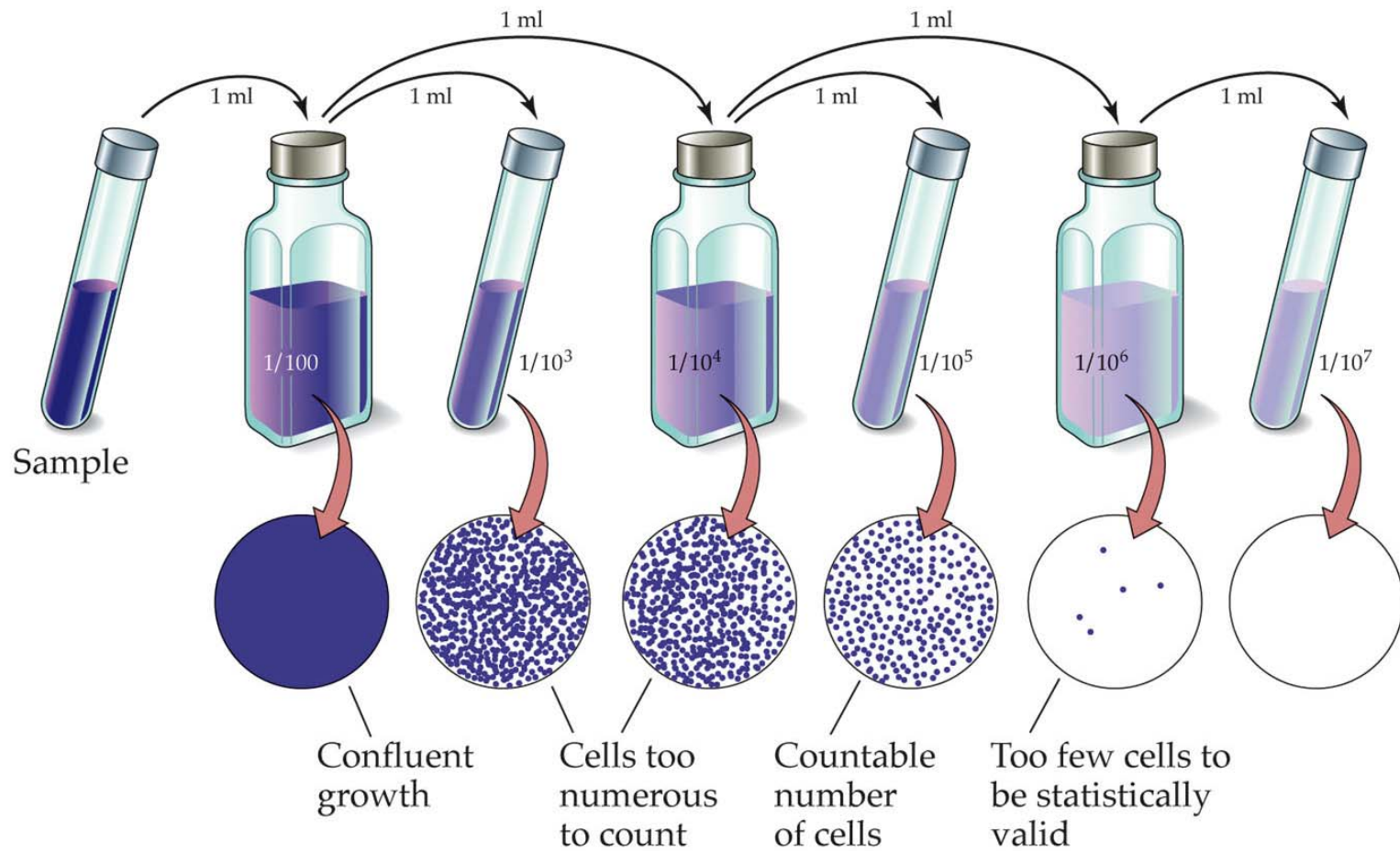
Sterile medium is added and
mixed well with inoculum

Incubation

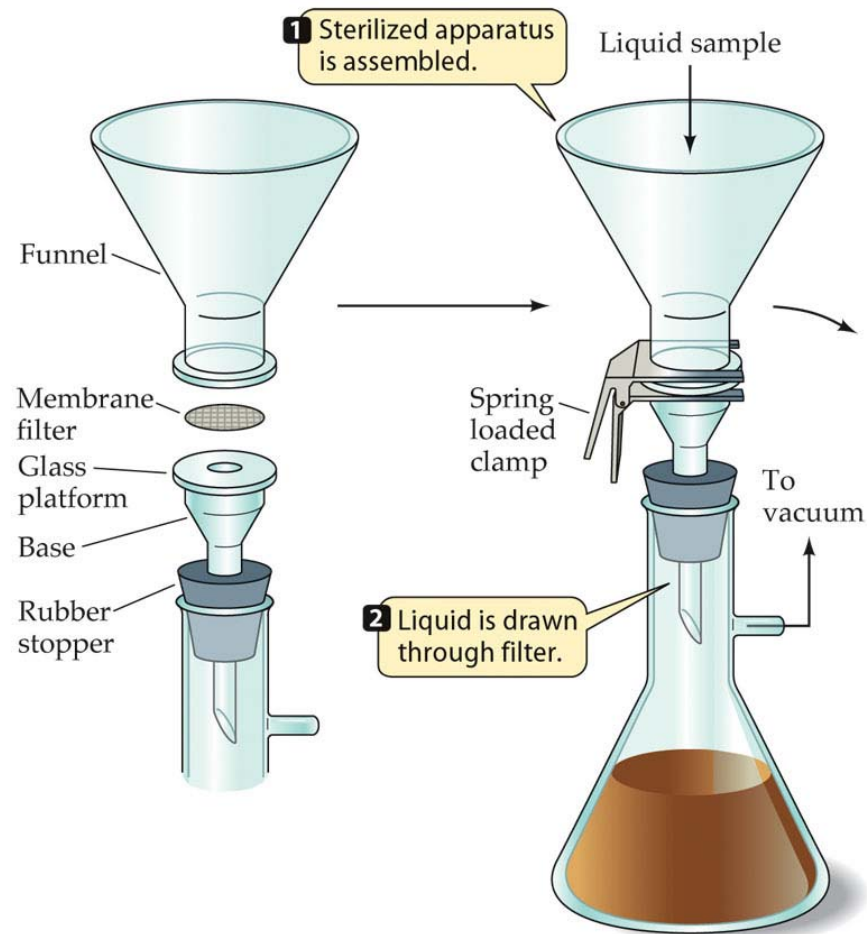


Typical pour-plate
results

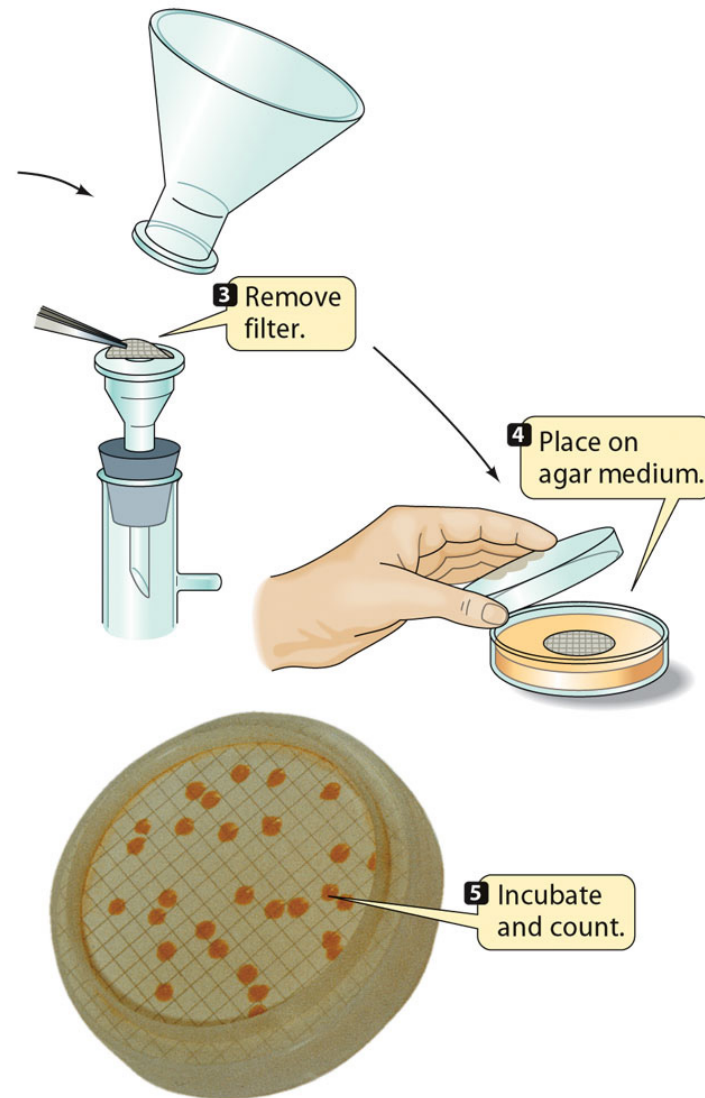
Counting the number of viable cells by serial dilution and plate count



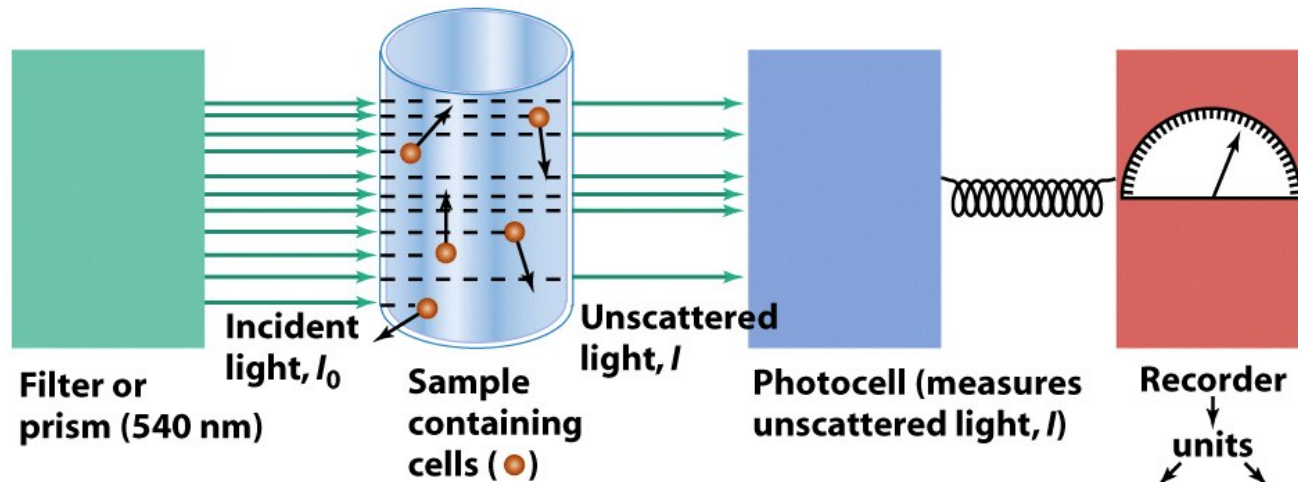
(Part 1) Concentration of cells by membrane filtration



(Part 2) Concentration of cells by membrane filtration



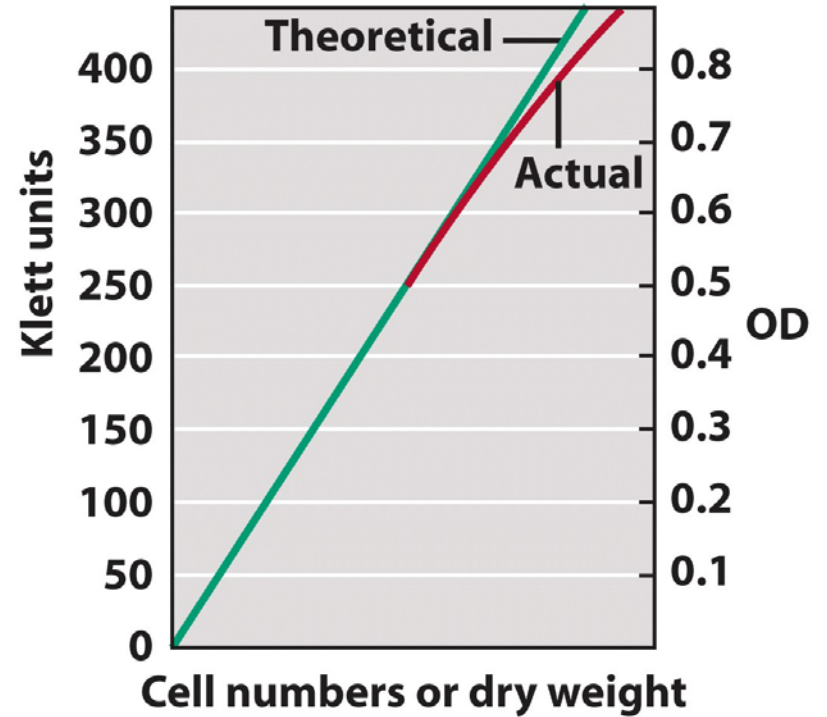
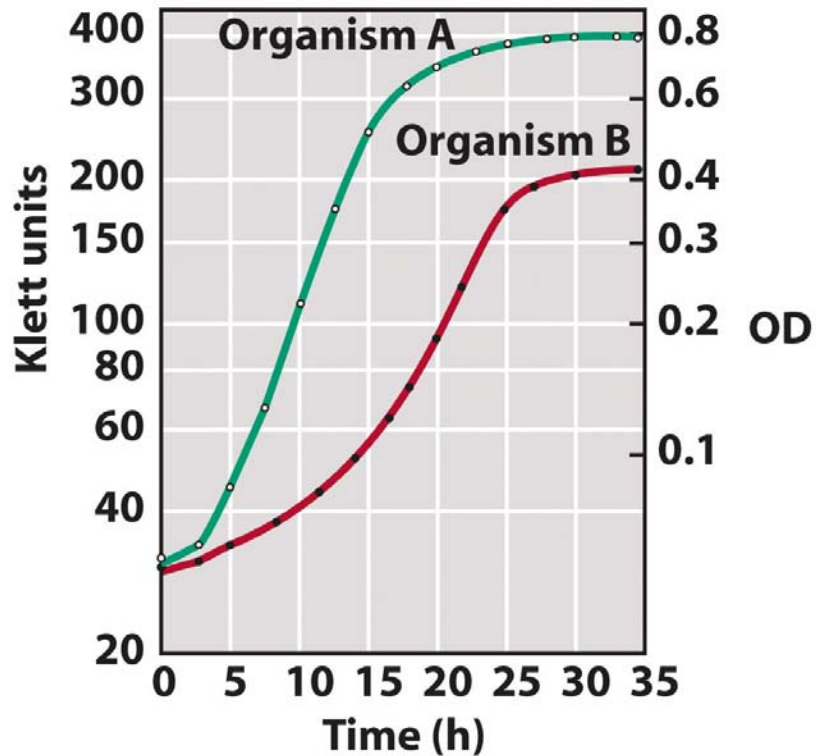
Turbidity measurements of microbial growth



Spectrophotometer—
optical density (OD)
 $= \text{Log } \frac{I_0}{I}$

Klett photometer—
Klett units $= \frac{\text{OD}}{0.002}$

Turbidity measurements of microbial growth

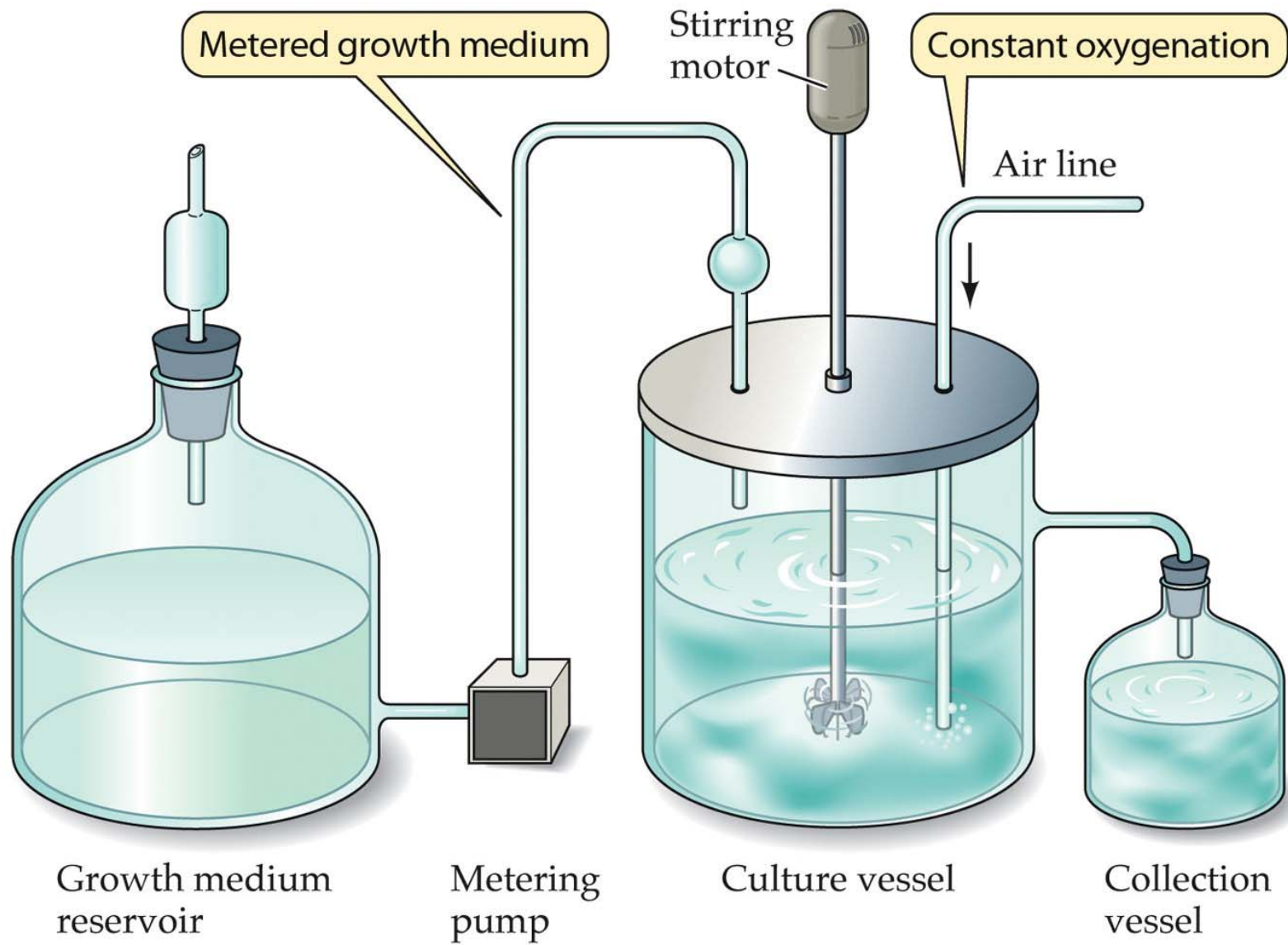


The Process of Growth

- Continuous Culture: The wonders of the **chemostat**
Steady State
Reproducible Physiology
Fine control

Key parameters: K_s , μ_{max} , Yield

Closed systems vs. Open systems vs. Nature!



Fresh medium
from reservoir

Sterile air or
other gas

Flow-rate
regulator

Gaseous
headspace

Culture
vessel

Culture

Overflow

Effluent containing
microbial cells

$$D = F/V = \mu$$

Where:

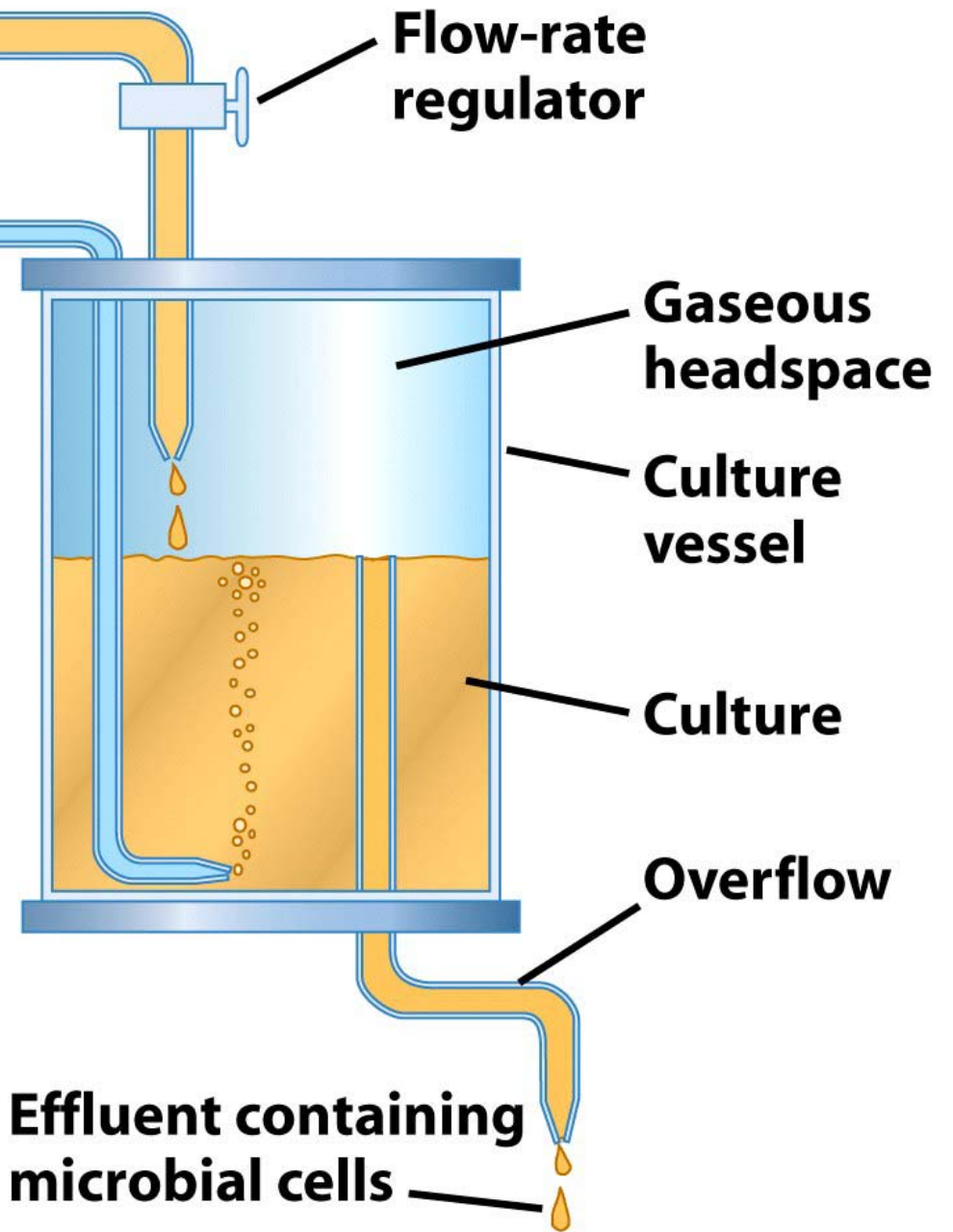
D = dilution rate

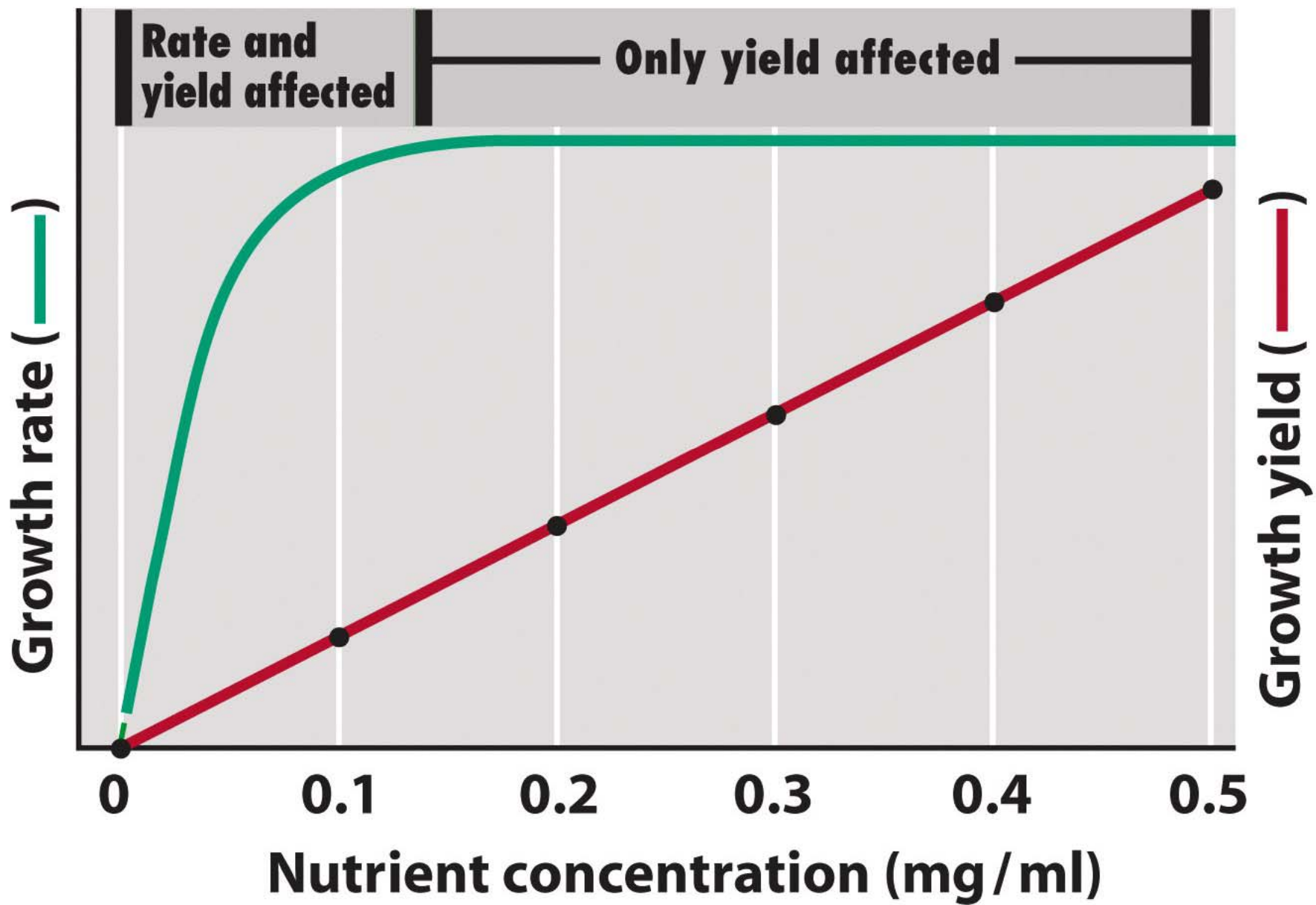
F = flow rate

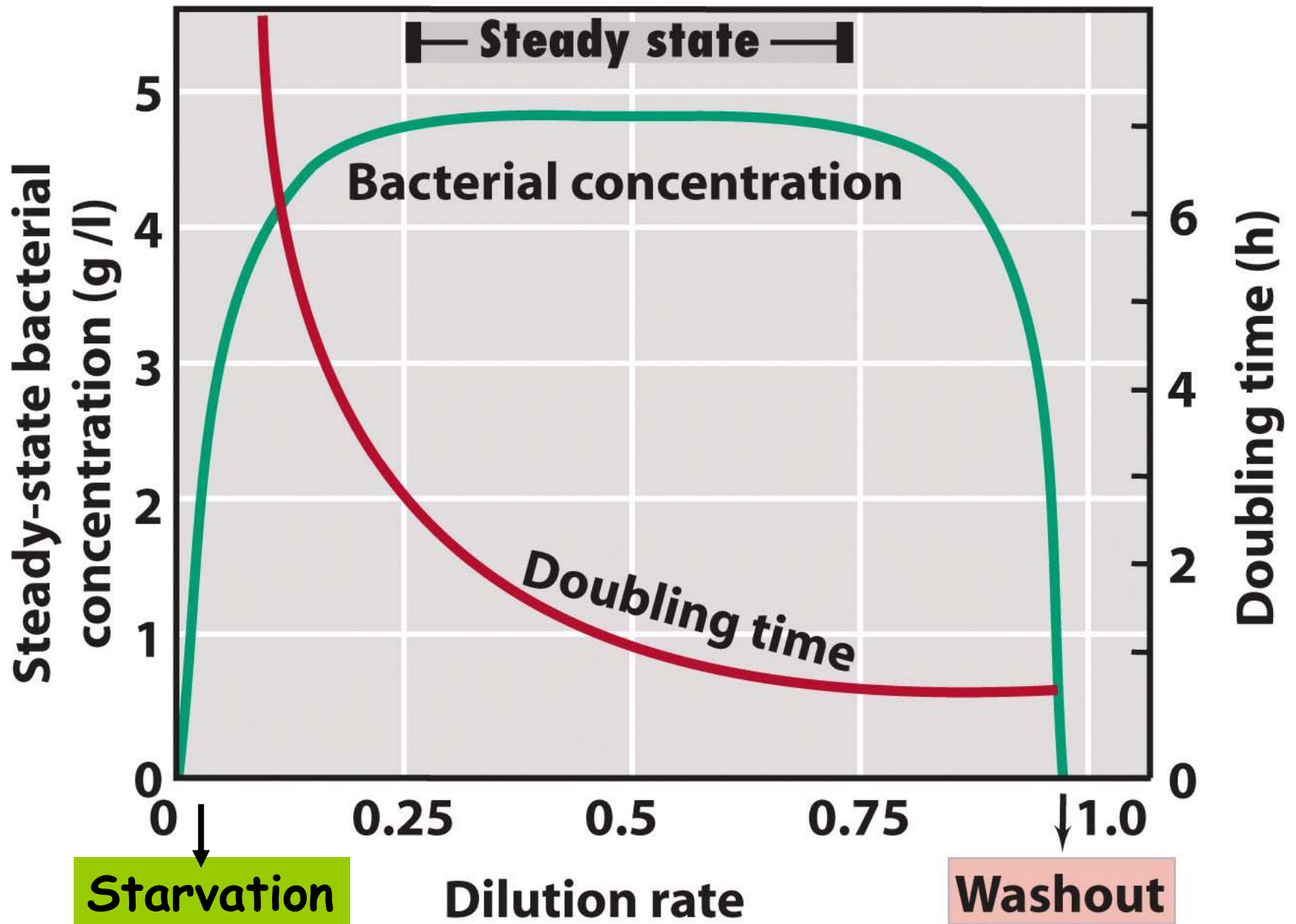
V = volume

μ = growth rate

Rem: At Steady State







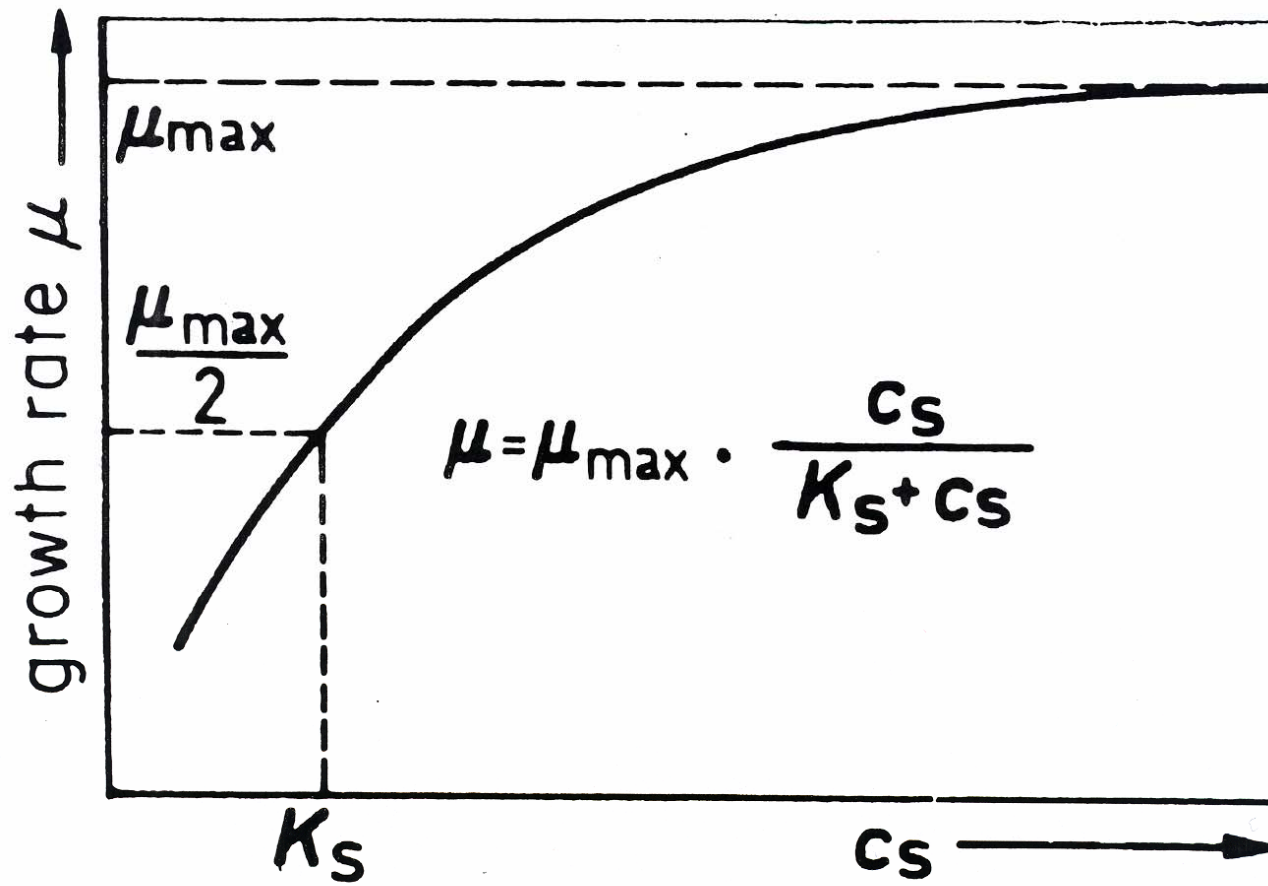


Fig. 6.10 *Dependence of growth rate μ on the substrate concentration c_s .*

Steady-state relationship between substrate concentration and output of bacterial mass

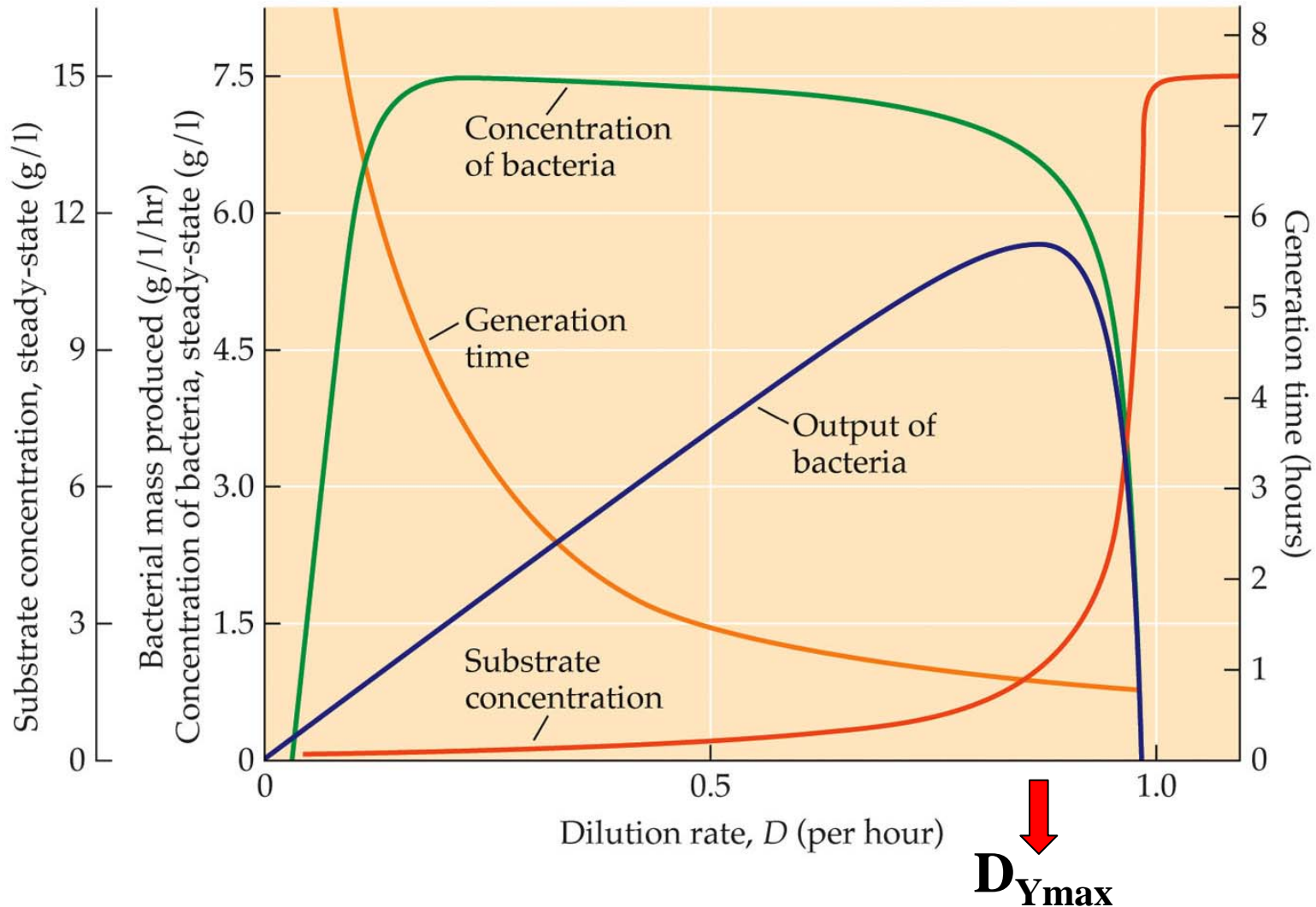


Table 6.2**Growth yields of anaerobic bacteria utilizing glucose as the energy source**

	Mol ATP/Mol Glucose	y_{\max} (g of cell/mol Glucose)	y_{ATP} (g of cell/mol ATP)
<i>Lactobacillus delbrueckii</i> ^a	2	21	10.5
<i>Enterococcus faecalis</i> ^a	2	20	10
<i>Zymomonas mobilis</i> ^b	1	9	9

^aHomolactic fermentation, Embden–Meyerhof pathway (see Chapter 10).

^bAlcoholic fermentation, Entner–Doudoroff pathway (see Chapter 10).